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Syringic acid, hepatoprotection, inflammation, hepatocellular carcinoma, apoptosis, hepG2, oxidative stress

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## **Syringic Acid and Liver Damage**

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## **ABSTRACT**

Data from valuable literature information on the impact of syringic acid (SA) values on health. Studies have succeeded in pointing out that with vanillic acid, SA members can suppress hepatic fibrosis in chronic artifacts and instead point to its potential to mitigate widespread damage. It is also impressive that it effectively alleviates SA ammonia penalties and thioacetamide-induced hepatic tattooing of commonly occurring tattoo markers such as AST, ALT, ALP and LDH. SA showed promise in alleviating diabetic diabetes complications and protecting against renal, neuronal, cardiac and hepatic damage in rats. Contrary to these positive effects, another study shows that SA can induce the effects shown in clinical trials and emphasizes that the side effects of its effects are spent and affect more.

## INTRODUCTION

Herbal metabolites are accepted as the source of natural products with different structures and biological effects. Recently, there has been a growing interest among researchers in the use of natural polyphenols as potential therapeutic phytochemicals due to their biological properties. Phenolic acids are the major polyphenols found in plants<sup>[1-5]</sup>.

Syringic acid (SA) has antioxidant<sup>[1]</sup>, antiinflammatory<sup>[2]</sup>, neuroprotective<sup>[3]</sup>, hepatoprotective<sup>[4]</sup>, cardioprotective<sup>[5]</sup>, nephroprotective and antiantidiabetic<sup>[8-10]</sup> properties<sup>[6,7]</sup>. The antiglycator<sup>[11]</sup>, antisteatosis-anti-inflammatory<sup>[12]</sup>, antihypertensive  $^{\left[13,14\right]}$  and antibacterial-antimic robial properties of SA have been reported<sup>[1,15]</sup>. Thanks to its neuroprotective [16-21] and hepatoprotective properties [22,23], it ameliorates diabetic cataracts by suppressing the aldose reductase enzyme. It has also been stated that SA is used to make dental cement<sup>[22]</sup>. SA reduces acute thromboembolism and clot formation in mice<sup>[24]</sup>. SA is a derivative of 4-hydroxy-3,5-dimethoxybenzoic acid and hydroxybenzoic acid (Fig. 1). It is found in different plants such as Herba dendrobii, Radix isatidis and Alpinia calcarata Roscoe<sup>[25-27]</sup>. SA has also been proven to protect the heart, liver and brain [4,6-9,23,28-29]. SA acts as a free radical scavenger and can fight against reactive oxygen species (ROS). It also has the potential to regulate enzyme activity, transcription factors, growth factors and signaling pathways<sup>[4]</sup>. In addition, no adverse effects were observed in toxicological studies with SA<sup>[30]</sup>.

The efficacy of antioxidants in reducing the risk of diabetic complications is remarkable<sup>[31,32]</sup>. Diabetes mellitus is a metabolic disease characterized by insufficient insulin secretion<sup>[33,34]</sup>. These pathological conditions can lead to irregular carbohydrate, protein and lipid metabolism<sup>[35]</sup>. The kidneys, eyes, nerves, liver, blood vessels and heart are the organs most affected by abnormal hyperglycemia<sup>[33]</sup>. The increase in diabetes, which is estimated to reach 53.1 million by 2025, is significant<sup>[36]</sup>. According to Rashedinia *et al.*<sup>[37]</sup>. They reported that dietary supplementation of SA

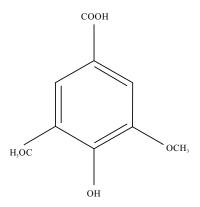


Fig. 1: Chemical structure of SA

protects hepatic tissue against hyperglycemia and lipid peroxidation. They also determined that SA provided protection by improving the deterioration in mitochondrial biogenesis in diabetic rats. The results of this study demonstrate the role of SA in mitochondrial biogenesis and its antioxidant activity in diabetes complications. It induces oxidative stress by increasing the level of reactive oxygen species (ROS) produced due to hyperglycemia, excessive mitochondrial activity and stimulation of the NF-kB signaling pathway in phagocytes. Oxidative stress plays a critical role in diabetic complications due to its adverse effects on vital biomacromolecules such as proteins, lipids and DNA<sup>[38]</sup>. These cellular damages can lead to cell death via apoptosis and necrosis<sup>[39]</sup>. The liver plays a central role in carbohydrate metabolism and maintenance of normal glucose levels. In diabetic conditions, chronic hyperglycemia, insulin resistance and decreased peripheral glucose uptake lead to an increase in lipogenesis as well as hepatic glucose output [40]. There is a strong correlation between diabetes and alteration in mitochondrial function. Decreased mitochondrial density, ATP production and decreased mitochondrial mRNA levels are other complications in diabetes and insulin resistance<sup>[41]</sup>. Mitochondrial biogenesis is a process affected by hyperglycemia and insulin resistance. Therefore, mitochondrial biogenesis is considered in the latest treatment strategies<sup>[42]</sup>.

Type 2 diabetes mellitus is a non-communicable metabolic disease characterized by severe and persistent hyperglycemia due to decreased insulin secretion and increased insulin resistance. Poor control of hyperglycemia in diabetic patients often results in severe microvascular (diabetic nephropathy, neuropathy and cardiomyopathy) and macrovascular (peripheral vascular disease and cerebrovascular disease) complications<sup>[43-45]</sup>. Ghule et al.<sup>[46]</sup> and Shang et al. [47] emphasized that diabetic nephropathy, manifested as microalbuminuria, impaired urinary creatinine clearance rate, renal hypertrophy and glomerular sclerosis, is the most common cause of end-stage renal disease worldwide. Painful diabetic neuropathy affects all types of peripheral nerves, affecting almost all systems and organs in the body, including the long somatosensory nerves in the extremities, where it often causes sensory loss<sup>[48]</sup>. In addition, diabetic patients are 2-5 times more likely to experience cardiac dysfunction than non-diabetic patients [49]. Various structural abnormalities that result in left ventricular hypertrophy, systolic and diastolic dysfunction, or a combination thereof, are the main manifestations of diabetic cardiomyopathy<sup>[50]</sup>. In addition, hyperglycemia causes liver dysfunction, a common secondary diabetic complication<sup>[51,52]</sup>. According to Mirza et al.[53] reported that SA reduced hyperglycemia, polydipsia, polyphagia, polyuria,

relative organ weight, cardiac hypertrophic indices, inflammatory markers, cell injury markers, glycosylated hemoglobin, histopathological score and oxidative stress. However, they found that it increased the Na/K ATPase activity. In conclusion, they found that SA could significantly reduce diabetic complications and renal, neuronal, cardiac and hepatic damage in streptozotocin-induced neonatal (nSTZ) diabetic rats. Hepatic encephalopathy (HE) is a complex syndrome with neuropsychiatric abnormalities associated with acute liver dysfunctions such as cirrhosis [54]. The incidence of cirrhosis increases with time and cirrhosis is the most common risk factor for HE, particularly in patients with alcoholic liver failure and portal hypertension<sup>[55]</sup>. HE affects brain function and includes a wide variety of symptoms, including altered cognitive consciousness, impairment, sensory abnormalities, altered motor activity and personality defects, as well as spatial memory dysfunction<sup>[56-68]</sup>. The most prominent current theory regarding the pathogenesis of HE is the accumulation of ammonia and other neurotoxic substances in the central nervous system due to impaired liver function<sup>[59]</sup>. The effects of ammonia on astrocytes in the brain are largely notable for its role in neuronal damage and eventual cognitive and motor symptoms<sup>[60]</sup>. In addition, as a potent free radical generator, ammonia induces free radical formation in astrocytes and impairs the antioxidant capacity of astrocytes<sup>[61-62]</sup>. Okkay et al.<sup>[63]</sup> Effectively attenuates thioacetamide-induced liver injury through reduction in SA ammonia, AST, ALT, ALP, LDH and reduction in oxidative stress (low MDA, ROS and increased SOD and GSH), as well as TNF- $\alpha$ , IL-1 $\beta$  and decreased inflammatory damage by suppressing NF-kB and increasing IL-10. It has also been stated that SA reduces the aggravating effects of thioacetamide. In conclusion, syringic acid has been reported to exert hepatoprotective and neuroprotective effects against hepatic encephalopathy by reducing hepatotoxicity biomarkers, suppressing hyperammonemia, as well as exerting antioxidant and anti-inflammatory effects.

Hepatocellular carcinoma is the second most common cause of cancer death in the world and its incidence has increased significantly worldwide over the past two decades. Its incidence is increasing and is an important public health problem<sup>[64]</sup>. Recently, the International Agency for Research on Cancer, a specialized cancer organization of the World Health Organization, found that hepatocellular carcinoma is now the second leading cause of death worldwide. As the causes of hepatocellular carcinoma; chronic alcohol consumption, hepatitis B and C virus infections, non-alcoholic fatty liver diseases, foods contaminated with aflatoxin B1, etc., takes place<sup>[65]</sup>. The most common treatment modalities in current practice for hepatocellular carcinoma are surgery, ablation and

liver transplantation<sup>[66]</sup>. Sorafenib, a mitogen-activated protein kinase pathway inhibitor, is currently the only therapeutic agent approved for systemic use in hepatocellular carcinoma patients, although it has several adverse effects including hyperbilirubinemia, hand and foot skin reactions and fatigue<sup>[67]</sup>. Despite advances in the diagnosis and treatment of hepatocellular carcinoma, its incidence and mortality continue to increase. Therefore, there is a need for effective treatment options with fewer or possibly no side effects for patients with advanced hepatocellular carcinoma. Research into the efficacy of plant-based drugs has received increasing attention due to their little or no side effects. It has been reported that natural bioactive substances alter the redox state and interfere with essential cellular functions such as cell cycle, apoptosis, inflammation, angiogenesis<sup>[68]</sup>.

Several studies have shown that plant-derived phytocompounds have a broad spectrum of biological activity, including anti-inflammatory, antioxidant, antimutagenic and anticancer properties<sup>[69]</sup>. There are studies showing that SA has a cytotoxic effect on the human HepG2 cell line and is a promising agent in anticancer research. According to Itoh et al. [70] reported that administration of syringing acid and vanillic acid significantly reduced transaminase activity on concanavalin A (ConA)-induced liver injury in mice. In addition, they stated that the administration of syringe acid and vanillic acid significantly suppressed cytokine levels. It has also been stated that SA treatment causes significant cytotoxicity and ROS release in HepG2 cells. Gheena and Ezhilarasan<sup>[71]</sup> reported that SA has a cytotoxic effect on the human HepG2 cell line and can be used as a promising agent in anticancer research.

Drug-induced liver injury is increasingly recognized as one of the leading causes of acute liver failure and the need for liver transplantation<sup>[72]</sup>. Drug-induced liver injury is one of the common reasons for withdrawal of drugs from the market and clinical trials. For example, acetaminophen, amoxicillin/clavulanate, dapsone, isoniazid and methotrexate are the most common causes of DILI<sup>[73-74]</sup>. Gheena *et al*.<sup>[75]</sup> reported that SA could be a promising herbal medicine that can prevent Sodium valproate-induced hepatotoxicity when administered together due to its potential anti-inflammatory effects.

Obesity, which is risk factors for nonalcoholic fatty liver disease, nonalcoholic steatohepatitis, diabetes and related pathologies, is associated with insulin resistance and adipose tissue distribution [76]. Ham et~al. [77] reported that SA decreased body weight, visceral fat mass, serum levels of leptin, TNF $\alpha$ , IFN $\gamma$ , IL-6 and MCP-1, insulin resistance, hepatic lipid content and early fibrosis, while increasing adiponectin circulation. They stated that SA has anti-obesity, anti-

inflammatory and anti-steatotic effects through regulation of lipid metabolic and inflammatory genes. They also stated that SA could be a new natural therapeutic agent for obesity or non-alcoholic liver disease.

In conclusion, the immunomodulatory properties of SA are also believed to play a role in the suppression of liver damage caused by the activation of T cells. Furthermore, a study conducted on rats fed a high-fat diet showed that SA has beneficial effects on dietinduced hepatic dysfunction and further supports its potential as a liver health protective agent. In general, studies have been conducted suggesting its promising effects in alleviating liver diseases and inflammation, as well as protecting against oxidative damage in SA diabetic cases. More research is needed to fully understand the mechanisms underlying these effects and to identify potential therapeutic applications of SA on liver health.

#### **REFERENCES**

- Shi, C., Y. Sun, Z. Zheng, X. Zhang and K. Song et al., 2016. Antimicrobial activity of syringic acid against cronobacter sakazakii and its effect on cell membrane. Food Chem., 197: 100-106.
- 2. Pezzuto, J.M., 2008. Grapes and human health: A perspective. J. Agric. Food Chem., 56: 6777-6784.
- 3. Pacheco-Palencia, L.A., S. Mertens-Talcott and S.T. Talcott, 2008. Chemical composition, antioxidant properties and thermal stability of a phytochemical enriched oil from Açai (*Euterpe oleracea* Mart.). J. Agric. Food Chem., 56: 4631-4636.
- Srinivasulu, C., M. Ramgopal, G. Ramanjaneyulu, C.M. Anuradha and C.S. Kumar, 2018. Syringic acid (SA): A review of its occurrence, biosynthesis, pharmacological and industrial importance. Biomed. Pharmacother., 108: 547-557.
- Ji, J., X. Yang, M. Flavel, Z.P.I. Shields and B. Kitchen, 2019. Antioxidant and anti-diabetic functions of a polyphenol-rich sugarcane extract. J. Am. Coll. Nutr., 38: 670-680.
- Güven, M., A.B. Aras, N. Topaloğlu, A. Ozkan and H.M. Sen *et al.*, 2015. The protective effect of syringic acid on ischemia injury in rat brain. Turk. J. Med. Sci., 45: 233-240.
- Yunhai, L., F. Jianguo, L. Ting, W. Wenqing and L. Aihua, 2003. Anti-endotoxic effects of syringic acid ofradix isatidis. J. Huazhong Univ. Sci. Technol. Med. Sci., 23: 206-208.
- Muthukumaran, J., S. Srinivasan, R.S. Venkatesan, V. Ramachandran and U. Muruganathan, 2013. Syringic acid, a novel natural phenolic acid, normalizes hyperglycemia with special reference to glycoprotein components in experimental diabetic rats. J. Acute Dis., 2: 304-309.

- Srinivasan, S., J. Muthukumaran, U. Muruganathan, R.S. Venkatesan and A.M. Jalaludeen, 2014. Antihyperglycemic effect of syringic acid on attenuating the key enzymes of carbohydrate metabolism in experimental diabetic rats. Biomed. Preventive Nutr., 4: 595-602.
- Pawar, S., A. Upaganlawar and C. Upasani, 2021. Evaluation of some phenolic acids in diabetic neuropathy. Indian J. Pharm. Educ. Res., 55: 176-183.
- 11. Bhattacherjee, A. and A. Datta, 2015. Mechanism of antiglycating properties of syringic and chlorogenic acids in in vitro glycation system. Food Res. Int., 77: 540-548.
- Li, Y., L. Zhang, X. Wang, wei wu and R. Qin, 2018. Effect of syringic acid on antioxidant biomarkers and associated inflammatory markers in mice model of asthma. Drug Dev. Res., 80: 253-261.
- Fernández, M.A., M.T. Sáenz and M.D. García, 1998. Anti-inflammatory activity in rats and mice of phenolic acids isolated from scrophularia frutescens. J. Pharm. Pharmacol., 50: 1183-1186.
- 14. Tokmak, M., Y. Yuksel, M.H. Sehitoglu, M. Guven and T. Akman *et al.*, 2015. The neuroprotective effect of syringic acid on spinal cord ischemia/reperfusion injury in rats. Inflammation, 38: 1969-1978.
- Tokmak, M., M.H. Sehitoglu, Y. Yuksel, M. Guven and T. Akman et al., 2015. The axon protective effects of syringic acid on ischemia/reperfusion injury in a rat sciatic nerve model. Turk. Neurosurg., 27: 124-132.
- Rashedinia, M., M. Alimohammadi, N. Shalfroushan, M.J. Khoshnoud, M. Mansourian, N. Azarpira and Z. Sabahi, 2020. Neuroprotective effect of syringic acid by modulation of oxidative stress and mitochondrial mass in diabetic rats. BioMed Res. Int., Vol. 2020, No. 12. 10.1155/2020/8297984
- Dalmagro, A.P., A. Camargo and A.L.B. Zeni, 2017.
  Morus nigra and its major phenolic, syringic acid, have antidepressant-like and neuroprotective effects in mice. Metab. Brain Dis., 32: 1963-1973.
- Cao, Y., L. Zhang, S. Sun, Z. Yi, X. Jiang and D. Jia, 2016. Neuroprotective effects of Syringic acid against ODG/R-induced injury in cultured hippocampal neuronal cells. Int. J. Mol. Med., 38: 567-573.
- 19. Zhao, Y., M. Dang, W. Zhang, Y. Lei, T. Ramesh, V.P. Veeraraghavan and X. Hou, 2020. Neuroprotective effects of syringic acid against aluminium chloride induced oxidative stress mediated neuroinflammation in rat model of Alzheimer's disease. J. Funct. Foods, Vol. 71. 10.1016/j.jff.2020.104009

- Ramachandran, V. and B. Raja, 2010. Protective effects of syringic acid against acetaminopheninduced hepatic damage in albino rats. J. Basic Clin. Physiol. Pharmacol., 21: 369-385.
- Itoh, A., K. Isoda, M. Kondoh, M. Kawase and A. Watari et al., 2010. Hepatoprotective effect of syringic acid and vanillic acid on CCl<sub>4</sub>-induced liver injury. Bio. Pharm. Bull., 33: 983-987.
- 22. Brauer, G.M. and J.W. Stansbury, 1984. Materials science cements containing syringic acid esters-oethoxybenzoic acid and zinc oxide. J. Dent. Res., 63: 137-140.
- 23. Manjunatha, S., A.H. Shaik, M.P.E., S.Y.A. Omar, A. Mohammad and L.D. Kodidhela, 2020. Combined cardio-protective ability of syringic acid and resveratrol against isoproterenol induced cardio-toxicity in rats via attenuating NF-kB and tnf-α pathways. Sci. Rep., Vol. 10, No. 1. 10.1038/s41598-020-59925-0
- 24. Choi, J.H. and S. Kim, 2018. Mechanisms of attenuation of clot formation and acute thromboembolism by syringic acid in mice. J. Funct. Foods, 43: 112-122.
- 25. Wei, X., D. Chen, Y. Yi, H. Qi and X. Gao et al., 2012. Syringic acid extracted from Herba dendrobii Prevents diabetic cataract pathogenesis by inhibiting aldose reductase activity. Evidence-Based Compl. Alt. Med., 2012: 1-13.
- Kong, W., Y. Zhao, L. Shan, X. Xiao and W. Guo, 2008. Thermochemical studies on the quantity–antibacterial effect relationship of four organic acids from *Radix Isatidis* on *Escherichia coli* growth Bio. Pharm. Bull., 31: 1301-1305.
- 27. Rahman, M.A. and M.S. Islam, 2015. Alpinia calcarata roscoe: A potential phytopharma cological source of natural medicine. Pharmacogn. Rev., 9: 55-62.
- 28. Kumar, S., P. Prahalathan and B. Raja, 2012. Syringic acid ameliorates L-NAME-induced hypertension by reducing oxidative stress. Naunyn-Schmiedeberg's Arch. Pharmacol., 385: 1175-1184.
- 29. Shahzad, S., S. Mateen, S.S. Naeem, K. Akhtar, W. Rizvi and S. Moin, 2019. Syringic acid protects from isoproterenol induced cardiotoxicity in rats. Eur. J. Pharmacol., 849: 135-145.
- Mirza, A.C. and S.S. Panchal, 2019. Safety evaluation of syringic acid: Subacute oral toxicity studies in wistar rats. Heliyon, Vol. 5, No. 8. 10.1016/j.heliyon.2019.e02129
- 31. Schaft, N.V., J.D. Schoufour, J. Nano, J.C.K.D. Jong and T. Muka *et al.*, 2019. Dietary antioxidant capacity and risk of type 2 diabetes mellitus, prediabetes and insulin resistance: The rotterdam study. Eur. J. Epidemiol., 34: 853-861.

- 32. Khoshnoud, M.J. Z. Sabahi, M. Moein, M. Rashedinia and S. Pourshahsavari, 2019. Attenuation of hyperlipidemia in diabetic and Triton x-100 induced hyperlipidemic rats by thymus daenensis celak extract. Trends Pharmacol. Sci., 5: 57-64.
- 33. Sabahi, Z., M.J. Khoshnood-Mansoorkhani, S.R. Namadi and M. Moein, 2016. Antidiabetic and synergistic effects study of anthocyanin fraction from berberis integerrima fruit on streptozotocin-induced diabetic rats model. Trends Phramaceutical Sci., 2: 43-50.
- Metwally, M.M.M., L.L.M. Ebraheim and A.A.A. Galal, 2018. Potential therapeutic role of melatonin on STZ-induced diabetic central neuropathy: A biochemical, histopathological, immunohistochemical and ultrastructural study. Acta Histochemica, 120: 828-836.
- Oboh, G., O.M. Agunloye, S.A. Adefegha, A.J. Akinyemi and A.O. Ademiluyi, 2015. Caffeic and chlorogenic acids inhibit key enzymes linked to type 2 diabetes (*in vitro*): A comparative study. J. Basic Clin. Physiol. Pharmacol., 26: 165-170.
- 36. Mantovani, A. and G. Targher, 2017. Type 2 diabetes mellitus and risk of hepatocellular carcinoma: Spotlight on nonalcoholic fatty liver disease. Ann. Transl. Med., 5: 270-270.
- Rashedinia, M., Z. Sabahi, M. Khoshnoud, B. Khalvati, S.S. Hashemi, Z.G. Farsani and H.M. Gerashi, 2020. Syringic acid improves oxidative stress and mitochondrial biogenesis in the liver of streptozotocin-induced diabetic rats. Asian Pac. J. Trop. Biomed., Vol. 10, No. 111. 10.4103/2221-1691.276317
- 38. Sadi, G., G. Şahin and A. Bostanci, 2018. Modulation of renal insulin signaling pathway and antioxidant enzymes with streptozotocin-induced diabetes: Effects of resveratrol. Medicina, Vol. 55, No. 1 .10.3390/medicina55010003
- 39. Pradeep, S.R. and K. Srinivasan, 2017. Amelioration of oxidative stress by dietary fenugreek (*Trigonella foenum-Graecum* L.) seeds is potentiated by onion (*Allium cepa* L.) in streptozotocin-induced diabetic rats. Applied Physiol., Nutr., Metab., 42: 816-828.
- 40. Gjorgjieva, M., G. Mithieux and F. Rajas, 2019. Hepatic stress associated with pathologies characterized by disturbed glucose production. Cell Stress, 3: 86-99.
- 41. Blake, R. and I.A. Trounce, 2014. Mitochondrial dysfunction and complications associated with diabetes. Biochim. Biophys. Acta (BBA) Gen. Subjects, 1840: 1404-1412.
- 42. Kim, J.A., Y. Wei and J.R. Sowers, 2008. Role of mitochondrial dysfunction in insulin resistance. Circulation Res., 102: 401-414.

- 43. Laakso, M., 2011. Heart in diabetes: A microvascular disease. Diabetes Care, 34: S145-S149
- 44. Brownlee, M., 2001. Biochemistry and molecular cell biology of diabetic complications. Nature, 414: 813-820.
- 45. OKAWA, H. and K. DOI, 1983. Neoplastic lesions in streptozotocin-treated rats. Exp. Anim., 32: 77-84.
- 46. Ghule, A.E., S.S. Jadhav and S.L. Bodhankar, 2012. Trigonelline ameliorates diabetic hypertensive nephropathy by suppression of oxidative stress in kidney and reduction in renal cell apoptosis and fibrosis in streptozotocin induced neonatal diabetic (nSTZ) rats. Int. Immunopharmacol., 14: 740-748.
- Shang, G., P. Gao, Z. Zhao, Q. Chen, T. Jiang, N. Zhang and H. Li, 2013. 3, 5-diiodo-l-thyronine ameliorates diabetic nephropathy in streptozotocin-induced diabetic rats. Biochim. Biophys. Acta (BBA) Mol. Basis Dis., 1832: 674-684.
- 48. .Albers, J.W. and R. Pop-Busui, 2014. Diabetic neuropathy: Mechanisms, emerging treatments and subtypes. Curr. Neurol. Neurosci. Rep., Vol. 14. 10.1007/s11910-014-0473-5
- 49. Herlitz, J., K. Malmberg, B.W. Karlson, L. Rydén and Å. Hjalmarson, 2009. Mortality and morbidity during a five-year follow-up of diabetics with myocardial infarction. Acta Med. Scand., 224: 31-38.
- 50. Hayat, S.A., B. Patel, R.S. Khattar and R.A. Malik, 2004. Diabetic cardiomyopathy: Mechanisms, diagnosis and treatment. Clin. Sci., 107: 539-557.
- 51. Rashid, K., J. Das and P.C. Sil, 2013. Taurine ameliorate alloxan induced oxidative stress and intrinsic apoptotic pathway in the hepatic tissue of diabetic rats. Food Chem. Toxicol., 51: 317-329.
- 52. Harrison, S.A., 2006. Liver disease in patients with diabetes mellitus. J. Clin. Gastroenterol., 40: 68-76.
- Mirza, A.C., S.S. Panchal, A.A. Allam, S.I. Othman, M. Satia and S.N. Mandhane, 2022. Syringic acid ameliorates cardiac, hepatic, renal and neuronal damage induced by chronic hyperglycaemia in wistar rats: A behavioural, biochemical and histological analysis. Molecules, Vol. 27, No. 19. 10.3390/molecules27196722
- 54. Pflugrad, H., A.B. Tryc, A. Goldbecker, H. Barg-Hock and C. Strassburg *et al.*, 2019. Cerebral metabolite alterations in patients with posttransplant encephalopathy after liver transplantation. PLOS ONE, Vol. 14, No. 8. 10.1371/journal.pone.0221626
- 55. Tapper, E.B., J.B. Henderson, N.D. Parikh, G.N. Ioannou and A.S. Lok, 2019. Incidence of and risk factors for hepatic encephalopathy in a population-based cohort of Americans with cirrhosis. Hepatology Commun., 3: 1510-1519.

- Erceg, S., P. Monfort, O. Cauli, C. Montoliu, M. Llansola, B. Piedrafita and V. Felipo, 2006. Role of extracellular cGMP and of hyperammonemia in the impairment of learning in rats with chronic hepatic failure. Neurochem. Int., 48: 441-446.
- McMillin, M., G. Frampton, M. Thompson, C. Galindo and H. Standeford et al., 2014. Neuronal CCL2 is upregulated during hepatic encephalopathy and contributes to microglia activation and neurological decline. J. Neuroinflammation, Vol. 11. 10.1186/1742-2094-11-121.
- 58. Gow, A.G., 2017. Hepatic encephalopathy. Vet. Clin. North Am.: Small Anim. Pract., 47: 585-599.
- 59. Felipo, V., 2013. Hepatic encephalopathy: Effects of liver failure on brain function. Nat. Rev. Neurosci., 14: 851-858.
- 60. Häussinger, D. and H. Sies, 2013. Hepatic encephalopathy: Clinical aspects and pathogenetic concept. Arch. Biochem. Biophys., 536: 97-100.
- 61. Germoush, M.O., S.I. Othman, M.A. Al-Qaraawi, H.M. Al-Harbi and O.E. Hussein *et al.*, 2018. Umbelliferone prevents oxidative stress, inflammation and hematological alterations and modulates glutamate-nitric oxide-cGMP signaling in hyperammonemic rats. Biomed. Pharmacother., 102: 392-402.
- 62. Hajipour, S., Y. Farbood, M. Dianat, M. Rashno, L.S. Khorsandi and A. Sarkaki, 2021. Thymoquinone improves behavioral and biochemical deficits in hepatic encephalopathy induced by thioacetamide in rats. Neurosci. Lett., Vol. 745. 10.1016/j.neulet.2020.135617
- Okkay, I.F., U. Okkay, O.L. Gundogdu, C. Bayram, A.S. Mendil, M.S. Ertugrul and A. Hacimuftuoglu, 2022. Syringic acid protects against thioacetamideinduced hepatic encephalopathy: Behavioral, biochemical and molecular evidence. Neurosci. Lett., Vol. 769. 10.1016/j.neulet.2021.136385
- 64. Hartke, J., M. Johnson and M. Ghabril, 2017. The diagnosis and treatment of hepatocellular carcinoma. Seminars Diagn. Pathol., 34: 153-159.
- Sanyal, A.J., S.K. Yoon and R. Lencioni, 2010.
  The etiology of hepatocellular carcinoma and consequences for treatment. Oncologist, 15: 14-22.
- 66. Mauer, K., R. O'Kelley, N. Podda, S. Flanagan and S. Gadani, 2015. New treatment modalities for hepatocellular cancer. Curr. Gastroenterol. Rep., Vol. 17, No. 19. 10.1007/s11894-015-0442-4
- 67. lavarone, M., G. Cabibbo, M. Biolato, C.D. Corte and M. Maida *et al.*, 2015. Predictors of survival in patients with advanced hepatocellular carcinoma who permanently discontinued sorafenib. Hepatology, 62: 784-791.

- Induja, M.P., D. Ezhilarasan and N.A. Vardhan, 2018. Evolvulus alsinoides methanolic extract triggers apoptosis in HepG2 cells. Avicenna J. Phytomed., 8: 504-512.
- 69. Miyata, T., 2007. Pharmacological basis of traditional medicines and health supplements as curatives. J. Pharmacol. Sci., 103: 127-131.
- Itoh, A., K. Isoda, M. Kondoh, M. Kawase, M. Kobayashi, M. Tamesada and K. Yagi, 2009. Hepatoprotective effect of syringic acid and vanillic acid on concanavalin A: induced liver injury. Bio. Pharm. Bull., 32: 1215-1219.
- 71. Gheena, S. and D. Ezhilarasan, 2019. Syringic acid triggers reactive oxygen species-mediated cytotoxicity in HepG2 cells. Hum. Exp. Toxicol., 38: 694-702.
- 72. Germani, G., S. Battistella, D. Ulinici, A. Zanetto and S. Shalaby *et al.*, 2021. Drug induced liver injury: From pathogenesis to liver transplantation. Minerva Gastroenterol., 67: 50-64.

- 73. Ezhilarasan, D., 2019. Dapsone-induced hepatic complications: It's time to think beyond methemoglobinemia. Drug Chem. Toxicol., 44: 330-333.
- 74. Ezhilarasan, D., 2021. Hepatotoxic potentials of methotrexate: Understanding the possible toxicological molecular mechanisms. Toxicology, Vol. 458. 10.1016/j.tox.2021.152840
- 75. Gheena, S., D. Ezhilarasan, K.S. Harini and S. Rajeshkumar, 2022. Syringic acid and silymarin concurrent administration inhibits sodium valproate induced liver injury in rats. Environ. Toxicol., 37: 2143-2152.
- Duarte, N., I.C. Coelho, R.S. Patarrão, J.I. Almeida, C. Penha-Gonçalves and M.P. Macedo, 2015. How inflammation impinges on NAFLD: A role for kupffer cells. BioMed Res. Int., Vol. 2015. 10.1155/2015/984578
- 77. Ham, J.R., H.I. Lee, R.Y. Choi, M.O. Sim, K.I. Seo and M.K. Lee, 2016. Anti-steatotic and anti-inflammatory roles of syringic acid in high-fat dietinduced obese mice. Food Funct., 7: 689-697.