

Dynamics of Cytokines Associated with IL-17 Producing Cells in Serum and Milk in Mastitis of Experimental Challenging with *Staphylococcus aureus* and *Escherichia coli* in Dairy Goats

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Abstract: Interleukin-17 is a critical pro-inflammatory cytokine in the development of autoimmunity and the immune responses against infection of bacteria, fungus and parasites. In the present study, dynamics of IL-17 and cytokines associated with IL-17 producing in serum and milk in experimental mastitis challenged with *S. aureus* and *E. coli* in dairy goats were monitored using commercial ELISA kits. The results showed that the levels of IL-17 in milk were peaked at 24 or 48 h post challenged with *E. coli* or *S. aureus*, respectively but no detectable peak was found in serum. The levels of TGF- β , IL-6 and IL-1 β in milk were elevated in goats challenged with *E. coli* or *S. aureus* but only slight fluctuant were found in serum. These indicated that IL-17 was an important cytokine in the inflammation development of dairy goat mastitis challenged with *E. coli* or *S. aureus* and the local pro-inflammatory cytokines milieu plays an important role in the development of subclinical mastitis whether infected with *E. coli* or *S. aureus*.

Key words: IL-17, mastitis inflammation, dairy goats, cytokines, China, ELISA kits

INTRODUCTION

Mastitis, a highly prevalent disease in dairy goats, cows and ewes is known to affect the production and quality of milk, animal health and even threat human health through consumption of milk (Leitner *et al.*, 2008; Halasa *et al.*, 2009; Le Marechal *et al.*, 2009, 2011). The majority of clinical and subclinical mastitis cases in goat were caused by *Staphylococcus aureus*, *Streptococcus uberis* and *Escherichia coli* (Pisoni *et al.*, 2010). Therefore, prevention and control procedures to mastitis of animals mainly depend on antibiotic therapy in dry period in clinic (McDougall *et al.*, 2010; Bradley *et al.*, 2011). However, considering the shortcomings of residues and resistant of antibiotics (Vaarst *et al.*, 2006; Virdis *et al.*, 2010), new non-antibiotic therapy strategies are expected to be developed. Recently, several studies showed that the detail understanding the defense mechanism of mammary glands and development of inflammation in mastitis are the key issues to control mastitis (Mazzilli and Zecconi, 2010; Rinaldi *et al.*, 2010).

Interleukin-17 (IL-17) is an important pro-inflammatory cytokine which plays a critical role in the development of inflammation in autoimmunity (allergic asthma, rheumatoid arthritis, systemic lupus

erythematosus inflammatory bowel disease) (Yen *et al.*, 2006; Doreau *et al.*, 2009; Shahrara *et al.*, 2009; Wang *et al.*, 2010) and immune responses against infection of bacterial (*Klebsiella pneumonia*, *S. aureus*, *listeria monocytogenes*) (Aujla *et al.*, 2008; Hamada *et al.*, 2008; Ishigame *et al.*, 2009), fungal (*Candida albicans*) (Huang *et al.*, 2004). In these processes, IL-17 could induce and enhance expression of several chemokines (CXCL1, CXCL8) which would indirectly activate and recruit neutrophils into the impaired tissue to trigger effective immune response (Park *et al.*, 2005; Hartupée *et al.*, 2007; Yu *et al.*, 2007; Roussel *et al.*, 2010). In IL-17-deficient mice, collagen-induced arthritis was significantly suppressed (Nakae *et al.*, 2003) and IL-17RA^{-/-} mice were more susceptible to infected *Porphyromonas gingivalis* and *Candida albicans* than wild mice (Huang *et al.*, 2004; Yu *et al.*, 2008). Tao and mallard had reported that IL-17 mRNA was up-regulated in *S. aureus* mastitis of cows and IL-17 was more highly expressed in milk somatic cells than blood mononuclear cells but the level of IL-17 protein was not determined in this study (Tao and Mallard, 2007).

In the present study, the mastitis model in Guanzhong dairy goat was established through experimentally challenged with the two main pathogens of

S. aureus or *E. coli*, respectively and dynamics of the levels of IL-17 and cytokines associated with IL-17 producing in serum and milk were monitored in the development of mastitis.

MATERIALS AND METHODS

Animals: Six healthy Guanzhong dairy goats in mid-lactation were selected that were negative for LMT detection (Lanzhou Veterinary Research Institute, Chinese Academy of Agricultural Sciences, Lanzhou, Gansu province, China) and bacteriological analysis in milk. The bacteriological analysis was performed according to previous research (Moroni *et al.*, 2005).

Preparation of bacteria: The organisms used were *E. coli* and *S. aureus* which were originally isolated from subclinical cases of dairy goat mastitis and had been used to establish mastitis model in dairy goat (Mo *et al.*, 2011). Prior to intramammary infection each strain was spread onto trypticase soy agar plates and incubated overnight at 37°C and then a bacterial colony was selected and inoculated with nutrient broth for 18 h at 37°C. After determining the concentration of the stock cultures based on the spread plate colony counts, the stock was diluted in Phosphate Buffered Saline (PBS) to a final concentration of 3×10^3 Colony Forming Unit mL^{-1} (CFU mL^{-1}) of *E. coli* or 3×10^2 CFU mL^{-1} of *S. aureus*.

Intramammary challenge with *E. coli* or *S. aureus*: The dairy goat was infected immediately following the morning milking by hand. Briefly, the right half udder of each goat was infused with 1 mL of prepared *E. coli* ($n = 3$) or *S. aureus* ($n = 3$) inoculum and the contralateral udder of each animal was infused with 1 mL of PBS alone. The blood samples from vena jugularis and milk samples were collected pre-infection (time = 0) and post-infection at 4, 8, 24, 48 and 72 h, respectively.

Whey and serum preparation: For the preparation of whey, milk samples were centrifuged at 3,000 rpm for 30 min and the fat layer was removed with a spatula. The middle translucent supernatant was collected and stored at -20°C for further study. For the preparation of serum, vein blood samples were collected, clotted at room temperature for 30 min and centrifuged at 1,500 rpm for 10 min. The supernatant was collected and stored at -20°C.

Enzyme-Linked Immunosorbent Assays (ELISAs): The levels of IL-17, IL-6, IL-1 β and TGF- β in whey and serum were determined by commercial ELISA kits (Quantikine M Goat ELISA kit, R and D Systems) according to the manufacturer's instructions.

Statistical analysis: Results were expressed as mean \pm standard error and statistical significance was analyzed by the Student's t-test (Graphpad Prism Version 5.0 for Windows, Graphpad Software Inc., San Diego, CA, USA) with $p < 0.05$ considered significant.

RESULTS AND DISCUSSION

In the present study, two mastitis models of dairy goat challenged with *E. coli* or *S. aureus* were established. The levels of IL-17 in milk were elevated for two models with peaks at 24 or 48 h post challenged with *E. coli* or *S. aureus*, respectively (Fig. 1a). But the level of IL-17 in serum was presented to be slightly fluctuant with no peak detectable from 0-72 h post challenged with *E. coli* or *S. aureus* (Fig. 1b).

Then the levels of cytokines associated with IL-17 producing were studied. The levels of IL-6 in serum and milk were elevated with peaks at 24 and 48 h post challenged with *E. coli* (Fig. 2a, b). But for goats challenged with *S. aureus*, the level of IL-6 was slightly elevated only in milk at 48 h post infection and no detectable increase was found in serum (Fig. 2a, b). Both the levels of IL-1 β and TGF- β in serum were presented to be slightly fluctuant from 0-72 h post challenged

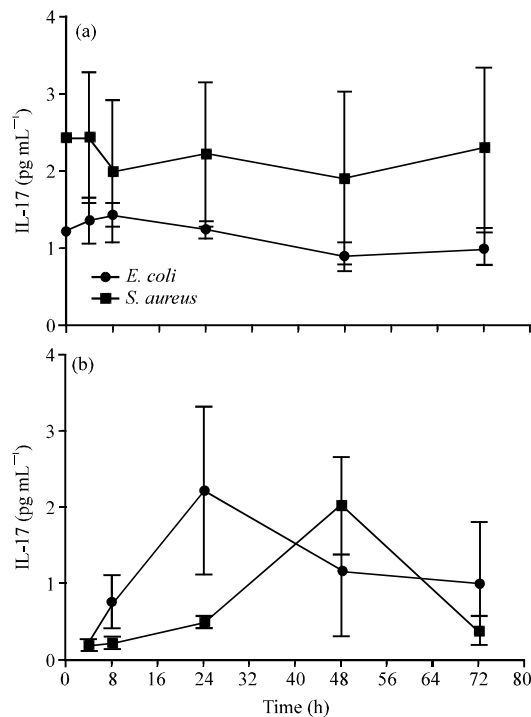


Fig. 1: The level of IL-17 in the blood (a) and milk (b) at different time points post challenged with *E. coli* or *S. aureus* in dairy goats

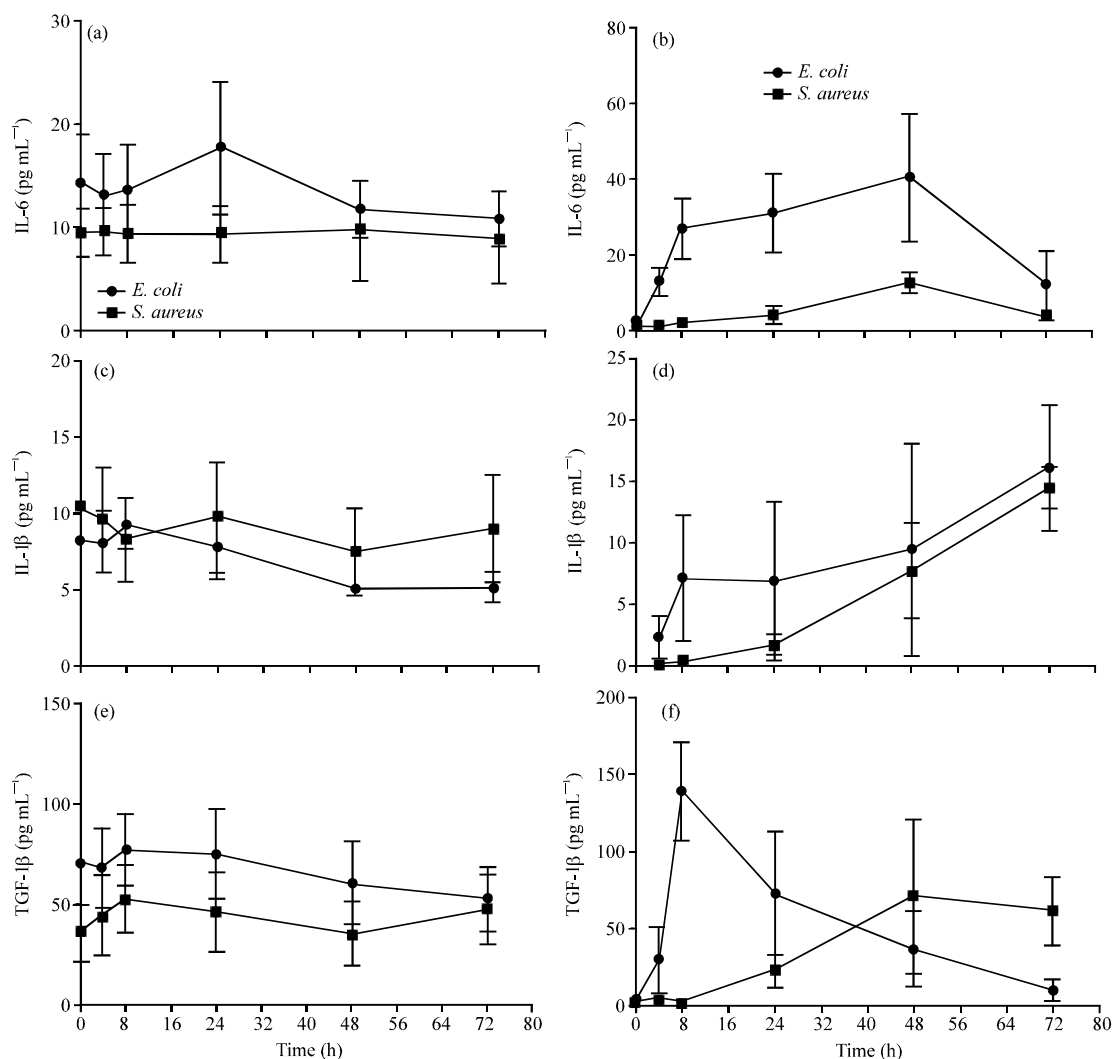


Fig. 2: The levels of IL-6 (a, b), IL-1β (c, d) and TGF-β (e, f) in the blood (a, c, e) and milk (b, d, f) at different time points post challenged with *E. coli* or *S. aureus* in dairy goats

with *S. aureus* and the levels of these two cytokines were decreased from 8-72 h post challenged with *E. coli* in serum (Fig. 2c, e).

However, the levels of IL-1β were gradually elevated from 8-72 h post challenged with *E. coli* and *S. aureus* in milk (Fig. 2d). The peak of TGF-β level was found in milk at 8 and 48 h post challenged with *E. coli* and *S. aureus*, respectively (Fig. 2f).

IL-17, a key pro-inflammatory cytokine, is produced by Th17 cells and several innate immune cells including γδ T cells, natural killer cells in a cytokine milieu (Roark *et al.*, 2008; Cua and Tato, 2010; Hemdan *et al.*, 2010) and play pleiotropic biological effects on multiple immune and non-immune cells (Tanaka *et al.*, 2009). The present study showed that the levels of IL-17 were

elevated in milk of goats challenged with *S. aureus* and *E. coli* which was in accordance with previous studies in mastitis of cows (Riollet *et al.*, 2006; Tao and Mallard, 2007).

Therefore, IL-17 would be an important cytokine in development of inflammatory response in mastitis of ruminants.

Local host immune response in mammary gland has been reported *in vivo* and *in vitro* (Beecher *et al.*, 2009; Moyes *et al.*, 2009, 2010). In the present study, both the changes in levels of IL-17 and IL-17 associated cytokines (TGF-β, IL-6 and IL-1β) in milk were more viable than that in serum. These results further confirmed that mastitis is only a local immune defense against pathogens in mammary gland.

CONCLUSION

IL-17 was an important cytokine in the inflammation development of dairy goat mastitis challenged with *E. coli* or *S. aureus*. However, the levels of IL-17 and cytokines associated with IL-17 producing were highly elevated in milk and slight fluctuant were detected in serum. These results indicated that the local pro-inflammatory cytokines milieu plays an important role in the development of subclinical mastitis whether infected with *E. coli* or *S. aureus* which should be considered in the therapy for mastitis.

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REFERENCES

- Aujla, S.J., Y.R. Chan, M. Zheng, M. Fei and D.J. Askew *et al.*, 2008. IL-22 mediates mucosal host defense against Gram-negative bacterial pneumonia. *Nat. Med.*, 14: 275-281.
- Beecher, C., M. Daly, D.P. Berry, K. Klostermann and J. Flynn *et al.*, 2009. Administration of a live culture of *Lactococcus lactis* DPC 3147 into the bovine mammary gland stimulates the local host immune response, particularly IL-1 α and IL-8 gene expression. *J. Dairy Res.*, 76: 340-348.
- Bradley, A.J., J.E. Breen, B. Payne and M.J. Green, 2011. A comparison of broad-spectrum and narrow-spectrum dry cow therapy used alone and in combination with a teat sealant. *J. Dairy Sci.*, 94: 692-704.
- Cua, D.J. and C.M. Tato, 2010. Innate IL-17-producing cells: The sentinels of the immune system. *Nat. Rev. Immunol.*, 10: 479-489.
- Doreau, A., A. Belot, J. Bastid, B. Riche and M.C. Trescol-Biemont *et al.*, 2009. Interleukin 17 acts in synergy with B cell-activating factor to influence B cell biology and the pathophysiology of systemic lupus erythematosus. *Nat. Immunol.*, 10: 778-785.
- Halasa, T., M. Nielen, R.B.M. Huirne and H. Hogeveen, 2009. Stochastic bio-economic model of bovine intramammary infection. *Livestock Sci.*, 124: 295-305.
- Hamada, S., M. Umemura, T. Shiono, K. Tanaka and A. Yahagi *et al.*, 2008. IL-17A produced by $\gamma\delta$ T cells plays a critical role in innate immunity against listeria monocytogenes infection in the liver. *J. Immunol.*, 181: 3456-3463.
- Hartupée, J., C. Liu, M. Novotny, X. Li and T. Hamilton, 2007. IL-17 enhances chemokine gene expression through mRNA stabilization. *J. Immunol.*, 179: 4135-4141.
- Hemdan, N.Y., G. Birkenmeier, G. Wichmann, A.M. Abu El-Saad and T. Krieger *et al.*, 2010. Interleukin-17-producing T helper cells in autoimmunity. *Autoimmunity Rev.*, 9: 785-792.
- Huang, W., L. Na, P.L. Fidel and P. Schwarzenberger, 2004. Requirement of interleukin-17A for systemic anti-*Candida albicans* host defense in mice. *J. Infect. Dis.*, 190: 624-631.
- Ishigame, H., S. Kakuta, T. Nagai, M. Kadoki and A. Nambu *et al.*, 2009. Differential roles of interleukin-17A and -17F in host defense against mucocutaneous bacterial infection and allergic responses. *Immunity*, 30: 108-119.
- Le Marechal, C., G. Jan, S. Even, J.A. McCulloch and V. Azevedo *et al.*, 2009. Development of serological proteome analysis of mastitis by *Staphylococcus aureus* in ewes. *J. Microbiol. Methods*, 79: 131-136.
- Le Marechal, C., R. Thiery, E. Vautor and Y. Le Loir, 2011. Mastitis impact on technological properties of milk and quality of milk products-a review. *Dairy Sci. Technol.*, 91: 247-282.
- Leitner, G., N. Silanikove and U. Merin, 2008. Estimate of milk and curd yield loss of sheep and goats with intramammary infection and its relation to somatic cell count. *Small Rumin. Res.*, 74: 221-225.
- Mazzilli, M. and A. Zecconi, 2010. Assessment of epithelial cells' immune and inflammatory response to *Staphylococcus aureus* when exposed to a macrolide. *J. Dairy Res.*, 77: 404-410.
- McDougall, S., K. Supre, S. de Vlieghe, F. Haesebrouck, H. Hussein, L. Clausen and C. Prosser, 2010. Diagnosis and treatment of subclinical mastitis in early lactation in dairy goats. *J. Dairy Sci.*, 93: 4710-4721.
- Mo, S., Y.S. Han, L.H. Lei and D.K. Chen, 2011. Establishment of mastitis model in guanzhong dairy goats. *Prog. Vet. Med.*, 32: 51-55.
- Moroni, P., G. Pisoni, G. Ruffo and P.J. Boettcher, 2005. Risk factors for intramammary infections and relationship with somatic-cell counts in Italian dairy goats. *Prev. Vet. Med.*, 69: 163-173.
- Moyes, K., J.K. Drackley, J.L. Salak-Johnson, D.E. Morin, J.C. Hope and J.J. Loores, 2009. Dietary-induced negative energy balance has minimal effects on innate immunity during a *Streptococcus uberis* mastitis challenge in dairy cows during midlactation. *J. Dairy Sci.*, 92: 4301-4316.

- Moyes, K.M., J.K. Drackley, D.E. Morin, S.L. Rodriguez-Zas, R.E. Everts, H.A. Lewin and J.J. Loo, 2010. Mammary gene expression profiles during an intramammary challenge reveal potential mechanisms linking negative energy balance with impaired immune response. *Physiol. Genomics*,
- Nakae, S., A. Nambu, K. Sudo and Y. Iwakura, 2003. Suppression of immune induction of collagen-induced arthritis in IL-17-deficient mice. *J. Immunol.*, 171: 6173-6177.
- Park, H., Z. Li, X.O. Yang, S.H. Chang and R. Nurieva *et al.*, 2005. A distinct lineage of CD4 T cells regulates tissue inflammation by producing interleukin 17. *Nat. Immunol.*, 6: 1133-1141.
- Pisoni, G., P. Moroni, S. Genini, A. Stella and P.J. Boettcher *et al.*, 2010. Differentially expressed genes associated with *Staphylococcus aureus* mastitis in dairy goats. *Vet. Immunol. Immunopathol.*, 135: 208-217.
- Rinaldi, M., R.W. Li, D.D. Bannerman, K.M. Daniels and C. Evock-Clover *et al.*, 2010. A sentinel function for teat tissues in dairy cows: Dominant innate immune response elements define early response to *E. coli* mastitis. *Funct. Integr. Genomics*, 10: 21-38.
- Riollet, C., D. Mutuel, M. Duonor-Cerutti and P. Rainard, 2006. Determination and characterization of bovine interleukin-17 cDNA. *J. Interferon Cytokine Res.*, 26: 141-149.
- Roark, C.L., P.L. Simonian, A.P. Fontenot, W.K. Born and R.L. O'Brien, 2008. $\Gamma\delta$ T cells: An important source of IL-17. *Curr. Opin. Immunol.*, 20: 353-357.
- Roussel, L., F. Houle, C. Chan, Y. Yao and J. Berube *et al.*, 2010. IL-17 promotes p38 MAPK-dependent endothelial activation enhancing neutrophil recruitment to sites of inflammation. *J. Immunol.*, 184: 4531-4537.
- Shahrara, S., S.R. Pickens, A. Dorfleutner and R.M. Pope, 2009. IL-17 induces monocyte migration in rheumatoid arthritis. *J. Immunol.*, 182: 3884-3891.
- Tanaka, S., T. Yoshimoto, T. Naka, S. Nakae, Y. Iwakura, D. Cua and M. Kubo, 2009. Natural occurring IL-17 producing T cells regulate the initial phase of neutrophil mediated airway responses. *J. Immunol.*, 183: 7523-7530.
- Tao, W. and B. Mallard, 2007. Differentially expressed genes associated with *Staphylococcus aureus* mastitis of Canadian *Holstein* cows. *Vet. Immunol. Immunopathol.*, 120: 201-211.
- Vaarst, M., T.W. Bennedsgaard, I. Klaas, T.B. Nissen, S.M. Thamsborg and S. Ostergaard, 2006. Development and daily management of an explicit strategy of nonuse of antimicrobial drugs in twelve danish organic dairy herds. *J. Dairy Sci.*, 89: 1842-1853.
- Viridis, S., C. Scarano, F. Cossu, V. Spanu, C. Spanu and E.P. de Santis, 2010. Antibiotic resistance in *Staphylococcus aureus* and coagulase negative staphylococci isolated from goats with subclinical mastitis. *Vet. Med. Int.*, 10.4061/2010/517060
- Wang, Y.H., K.S. Voo, B. Liu, C.Y. Chen and B. Uygungil *et al.*, 2010. A novel subset of CD4⁺ T_H2 memory/effector cells that produce inflammatory IL-17 cytokine and promote the exacerbation of chronic allergic asthma. *J. Exp. Med.*, 207: 2479-2491.
- Yen, D., J. Cheung, H. Scheerens, F. Poulet and T. McClanahan *et al.*, 2006. IL-23 is essential for T cell-mediated colitis and promotes inflammation via IL-17 and IL-6. *J. Clin. Invest.*, 116: 1310-1316.
- Yu, J.J., M.J. Ruddy, G.C. Wong, C. Sfintescu and P.J. Baker *et al.*, 2007. An essential role for IL-17 in preventing pathogen-initiated bone destruction: recruitment of neutrophils to inflamed bone requires IL-17 receptor-dependent signals. *Blood*, 109: 3794-3802.
- Yu, J.J., M.J. Ruddy, H.R. Conti, K. Boonnanantanasarn and S.L. Gaffen, 2008. The interleukin-17 receptor plays a gender-dependent role in host protection against *Porphyromonas gingivalis*-induced periodontal bone loss. *Infect. Immunity*, 76: 4206-4213.