

## Polymorphism of Ovar-DRB1 Second Exon with PCR-RFLP Technique in Arabi Sheep Population of Khuzestan Province

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**Abstract:** Major Histocompatibility Complex (MHC) plays a pivotal role in antigen presentation for eliciting immune responses against invading pathogens. The best-characterized genetic control of disease resistance and immune response in animals are associated with MHC. Therefore in this study polymorphism of ovar-DRB1 in DNA samples of 111 heads Arabi sheep using Polymerase Chain Reaction (PCR) and Restriction Fragment Length Polymorphism (RFLP) method was examined. Identities of 18 different patterns and 8 distinct DRB1 alleles among Arabi sheep have been determined. There is not deviation in *Ovar-DRB1* gene from Hardy-Weinberg equilibrium in this population results showed. Obtained results have revealed that the exon 2 region of ovar-DRB1 was highly polymorphic on Arabi sheep.

**Key words:** Arabi sheep, MHC, ovar-DRB1, PCR-RFLP, Khuzestan province, Iran

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### INTRODUCTION

The Major Histocompatibility Complex (MHC) plays a central role in the immune response of vertebrates. The extreme polymorphism enables the host to recognize enormous numbers in *MHC* genes of foreign peptides to trigger an immune reaction. The MHC of sheep is known as ovar, located on chromosome 20 (Dukkipati *et al.*, 2006). Ovar-MHC is poorly characterised compared to other domestic animals.

However, its basic structure is similar to that of other mammals, comprising class I-III regions. The class I region is poorly characterised and there is a significant controversy over the number of classical class I loci. Initial studies in this regard relied mainly on the use of alloantisera in micro-lymphocytotoxicity assays. The class II region is better characterized but the class III region is the least. Products of the class I and II genes, MHC molecules play a pivotal role in antigen presentation required for eliciting immune responses against invading pathogens.

However when MHCs of different mammals are compared, some regions appear to be well conserved and others vary widely (Kelley *et al.*, 2005). In general, the class II and class III regions are orthologous, i.e., they are clearly derived from a single ancestor without being subjected to major rearrangements (except in ruminants)

and their gene order is conserved. Chardon *et al.* (1985) reported that the serologically detected class I sheep leukocyte antigens are coded by *MHC* genes. Class II genes are members of the immunoglobulin superfamily of genes which are functionally specialised for presentation of antigenic peptides mainly derived from extracellular proteins and parasites to the T cells receptors. The ovar class II genes encode polymorphic glycoprotein composed of non-covalently linked  $\alpha$  and  $\beta$ -subunits that play a basal role in the initiation of the immune response to pathogen-derived peptide antigens. The highly polymorphic genes of the MHC play a central role in the immune recognition of pathogens and parasites. These genes encode MHC class I and II molecules that present peptide antigens to T cells initiating immune recognition of foreign antigens (Falk *et al.*, 1991).

Andersson and Rask (1988) demonstrated that the most polymorphic among the *MHC* genes is the DRB locus. *Ovar-DRB* genes exist in multiple copies, some functional and others non-functional. Konnai *et al.* (2003) reported that 106 ovar-DRB1 alleles have been identified by DNA sequencing of exon 2 from various breeds of sheep. Several studies (Thursz *et al.*, 1997; Carrington *et al.*, 1999; Penn *et al.*, 2002; Stear *et al.*, 2005) have confirmed this selective advantage of MHC heterozygosity against infectious agents. The Arabi sheep is one of the most important Iranian native breeds.

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These animals are fat-tailed, medium-sized and well adapted to extreme temperatures and conditions in Khuzestan province and Southwestern part of Iran (Ezzatpour, 2003). The current study was aimed at analyze the genetic diversity of the Ovar-DRB1 locus in Arabi sheep by PCR-RFLP Technique.

## MATERIALS AND METHODS

**Animal and DNA extraction:** This study was conducted on a total of 110 heads Arabi sheep from four different regions of Khuzestan province as follows: Shadegan region (n = 30), Hoveyzeh region (n = 28), Dasht e Azadegan region (n = 27) and Mollasani region (n = 26). All the animals were unrelated and selected at random. From each animal about 3-5 cc of blood was collected from the jugular vein with vacuum tubes coated with EDTA and transported in the Central Laboratory of Agricultural and Natural Resources Ramin (Khuzestan) University and stored at -20°C until DNA extraction. Genomic DNA extracted from 100 µL of blood according to Boom *et al.* (1990) Method. Quality and quantity of DNA were measured by electrophoresis in agarose gel 0.8%.

**PCR-RFLP:** A region exon 2 of the *Ovar-DRB1* gene was amplified to produce a 296 bp fragment. Study of Konnai *et al.* (2003) were used and they were as follow sequencing: OLA-ERB1 (GC) 5'-CCG GAA TTC CCG TCT CTG CAG CAC ATT TCT T-3' and OLA-XRBI (5'-AGC TCG AGC GCT GCA CAG TGA AAC TC-3'). A total of 25 µL reaction mixture containing 100-150 ng DNA template, 15 pm of each primer, 200 µL of each dNTP, 1.5 mM MgCl<sub>2</sub>, 1 U Taq DNA polymerase and 2.5 mm PCR buffer 1x. The optimal follow-up reactions and the patterns of Thermal Gradient Method with 30 cycles under the most ideal conditions for proliferation area were selected.

At the first, starting denaturation (95°C for 5 min), the next step with the temperature 95°C for 30 sec and the temperature annealing each cycle, 64-65°C for 30 sec and 72°C for 1 min temperature was extension. The temperature of extension was 72°C in 10 min and then it was 4°C in 20 min in the final stage. Samples were resolved on 1.2% agarose gel electrophoresis.

After amplification of this gene polymorphism to the desired gene fragments by enzymatic digestion with enzymes were introduced in the researches. Enzyme RsaI, a four nucleotide sequence GGCC at the junction of C and G are identified and broken. After adding the final mix-up micro tube and centrifuged, the samples for 16 h at 37°C

was placed in a warm bath. Then 8 mL of digestion products by 2.5% agarose gel electrophoresis was used to determine the size bands 50 bp size markers. The amplified fragment were digested with RsaI restriction endonuclease and then subjected to electrophoretic separation in ethidium bromide-stained 2.5% agarose gel. Samples were electrophoresed for 90 min at 80 V. Then gels were photographed under UV light with a Gel\_Doc system. Investigation of allelic and genetic frequencies performed by Chi-square ( $\chi^2$ ) and Hardy-Weinberg equilibrium carried out by GenAlex 6.3 (Peakal and Smouse, 2006).

## RESULTS AND DISCUSSION

Ovar-DRB1 polymorphism was analyzed by PCR amplification and RFLP of Ovar-DRB1 exon 2 in Arabi sheep population in Khuzestan province. DNA bands of expected size (296 bp) of the following replication PCR were obtained (Fig. 1). When each amplified product was cleaved by restriction enzyme, 8 different patterns were observed in RsaI digests (Table 1, Fig. 2).

The RsaI restriction showed in study the 8 restriction patterns a-h. In Arabi sheep 6 homozygous genotypes: bb, cc, dd, ee, gg, hh and 12 heterozygous genotypes: ab, ac, bd, be, bf, bh, cd, ce, ch, de, dh, he were observed. Allele c and b had the highest allelic frequency (24.77 and 21.6%, respectively) and genotype ce and bh were the most frequent patterns exhibited (18 and 17.1%, respectively) in population of Arabi sheep. The number of alleles (na), expected alleles (ne), observed heterozygosity and expected heterozygosity rates were 8, 5.223, 0.667 and 0.809, respectively. Chi-square ( $\chi^2$ ) test indicated that this population was at Hardy-Weinberg equilibrium and any selection done for these loci in mentioned population.

Evaluation of level heterozygosity dependent on *MHC* gene in population is very high. That is explanatory of good genetic diversity because heterozygosity is one of importance indices in detect of the genetic diversity and always is polestar of animal's breeders.

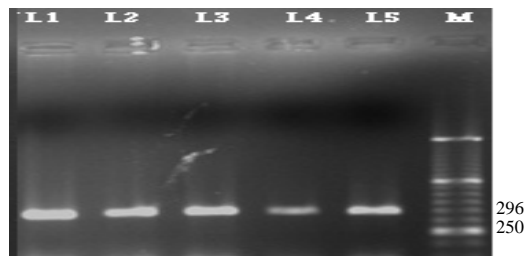
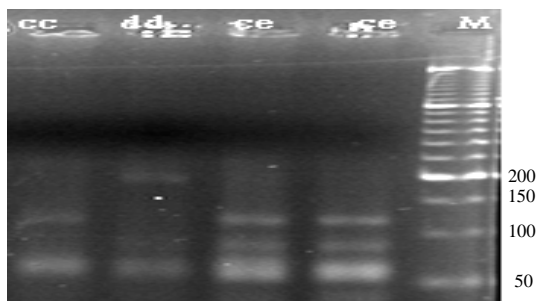


Fig. 1: 296 bp amplified fragment of *Ovar-MHC* genes

**Table 1: Allelic frequencies of the exon 2 of *MHC-DRB1* gene in Arabi sheep**

Alleles	Size of bands in identified alleles	Allelic frequency
A	296 bp	3.60
B	240/56 bp	21.60
C	117/69/56/54 bp	24.75
D	186/56/54 bp	9.90
E	84/69/56/54/33 bp	17.60
F	84/56/54/39/33/30 bp	1.35
G	84/56/54/39/30 bp	0.90
H	147/56/54/39 bp	20.30

Konnai *et al.* (2003)



**Fig. 2: PCR-RFLP analysis of the second exon of *Ovar-DRB1* gene (296 bp) from Arabi sheep**

## CONCLUSION

The findings of the present study confirmed and revealed that the exon 2 region of *Ovar-DRB1* was highly polymorphic in Arabi sheep and this population was at Hardy-Weinberg equilibrium too.

## REFERENCES

Andersson, L. and L. Rask, 1988. Characterization of the MHC class II region in cattle. The number of DQ genes varies between haplotypes. *Immunogenetics*, 27: 110-120.

Boom, R., C.J.A. Sol, M.M.M. Salimans, C.J. Jansen, P.M.E.W. van Dillen and J. van der Noordaa, 1990. Rapid and simple method for purification nucleic acid. *J. Clin. Microbiol.*, 28: 495-503.

Carrington, M., G.W. Nelson, M.P. Martin, T. Kissner and D. Vlahov *et al.*, 1999. HLA and HIV-1: Heterozygote advantage and B\*35-Cw\*04 disadvantage. *Science*, 283: 1748-1752.

Chardon, P., M. Kirszenbaum, P.R. Cullen, C. Geffrotin and C. Auffray *et al.*, 1985. Analysis of the sheep MHC using HLA class I, II and C4 cDNA probes. *Immunogenetics*, 22: 349-358.

Dukkipati, V.S.R., H.T. Blair, D.J. Garrick and A. Murray, 2006. Ovar-Mhc-ovine major histocompatibility complex: Structure and gene polymorphisms. *Genet. Mol. Res.*, 5: 581-608.

Ezzatpour, M., 2003. Native Iranian sheep and goat production. Moallef Publication, Tehran, Iran.

Falk, K., O. Rotzschke, S. Stevanovic, G. Jung and H.G. Rammensee, 1991. Allele-specific motifs revealed by sequencing of self-peptides eluted from MHC molecules. *Nature*, 23: 290-296.

Kelley, J., L. Walter and J. Trowsdale, 2005. Comparative genomics of major histocompatibility complexes. *Immunogenetics*, 56: 683-695.

Konnai, S., Y. Nagaoka, S. Takesima, M. Onuma and Y. Aida, 2003. Technical note: DNA typing for ovine MHC DRB1 using polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP). *J. Dairy Sci.*, 86: 3362-3365.

Peakal, R. and P.E. Smouse, 2006. GENALEX 6: Genetic Analysis in Excel. Population genetic software for teaching and research. *Mol. Ecol. Notes.*, 6: 288-295.

Penn, D.J., K. Damjanovich and W.K. Potts, 2002. MHC heterozygosity confers a selective advantage against multiple-strain infections. *Proc. Natl. Acad. Sci. USA.*, 99: 11260-11264.

Stear, M.J., G.T. Innocent and J. Buitkamp, 2005. The evolution and maintenance of polymorphism in the major histocompatibility complex. *Vet. Immunol. Immunopathol.*, 108: 53-57.

Thursz, M.R., H.C. Thomas, B.M. Greenwood and A.V.S. Hill, 1997. Heterozygote advantage for HLA class-II type in hepatitis B virus infection. *Nat. Genet.*, 17: 11-12.