ISSN: 1680-5593

© Medwell Journals, 2012

Measurement of Zinc in Arabian Horse's Serum by Spectrophotometric Method in Tabriz

¹Amirparviz Rezaeisaber, ²Reza Zarei and ³Mehrdad Nazeri ¹Department of Clinical Science, ²Department of Veterinary Medicine, ³Department of Veterinary Medicine, Tabriz Branch, Islamic Azad University, Tabriz, Iran

Abstract: Zinc is an essential mineral that is naturally present in some foods, added to others and available as a dietary supplement. Zinc is also found in many cold lozenges and some over-the-counter drugs sold as cold remedies. Zinc is involved in numerous aspects of cellular metabolism. It is required for the catalytic activity of approximately 100 enzymes and it plays a role in immune function, protein synthesis, wound healing, DNA synthesis and cell division. Zinc also supports normal growth and development during pregnancy, childhood and adolescence and is required for proper sense of taste and smell. A daily intake of zinc is required to maintain a steady state because the body has no specialized zinc storage system. The aim of this study was to measurement of zinc in Arabian horse's serum by Spectrophotometric Method in Tabriz. In this study about 186 blood samples from apparently healthy Arabian horses were obtained and blood samples taken near the ice and sent to the laboratory and after serum preparation were freezing inside the micro tube. In lab, blood samples centrifuged at 2000 rpm for 15 min.

Key words: Zinc, Arabian horses, serum, spectrophotometry, Tabriz, Iran

INTRODUCTION

Zinc is a trace element of essential biological importance. Zinc serves as structural ions in transcription factors and is stored and transferred in metallothionein, it is found in nearly 200 metalloenzymatic systems or zinc containing proteins. Three basic functions of relevance of this study have been demonstrated: catalytic, structural and as regulator for keratinocytes proliferation and differentiation, Marycz found morphologic improvement of hair after feeding with zinc enriched food.

dehydrogenase Alcohol possess antioxidant properties and also helps speed up wound healing and zinc deficiency has been shown to play a role in increased susceptibility to infections and delayed wound healing Wound healing of the skin and the cell metabolism both depend on zinc as a catalyst enzyme in DNA synthesis and in metallothionein mRNA. Zinc indirectly activates part of a cellular differentiation process: the keratinization that transforms live epithelial cells into corneous cells that are structurally stable and with no metabolic activity (Iwata et al., 1999). Pories et al. (1967) found that zinc sulphate significantly accelerates wound healing. In proteins, Brandt et al. (2009) studied the structural site of horse liver alcohol dehydrogenase and found that zinc ions often were coordinated to the amino acid side chains

of aspartic acid, glutamic acid, cysteine and histidine. Zinc ions are effective antimicrobial agents even at low concentrations (McCarthy et al., 1992). Studies of zinc deficiency including hair zinc levels have been reported in man and normal zinc status trough hair analysis seems potentially useful in experimental medicine but its use in clinical medicine will remain limited until validation by the standard methods of clinical investigation is achieved (Klevay et al., 1987). Clinical zinc deficiency has not been unequivocally described so far in the horse (Kienzle and Zorn, 2006) but several studies have been conducted in order to quantify the normal mineral status in orses. Zinc concentration in hair and serum are so far the most well studied paraclinical parameters to equine zinc status.

MATERIALS AND METHODS

This study was conducted to measurement of zinc in serum of Arabian horses during two seasons of 1 year Summer and Winter. In this study, about 186 blood samples from apparently healthy Arabian horses were obtained and blood samples taken near the ice and sent to the laboratory and after serum preparation were freezing inside the micro tube. In this inspection, age, body condition score and pregnancy status of animals was investigated. Simultaneous inspection of animals,

attempting to obtain blood samples of 10 mL of jugular vein was done by venoject. In lab, blood samples centrifuged at 2000 RPM for 15 min. Measurement of zinc in serum was followed by Spectrophotometric Method. In this study to analyzing and comparison of data were used of ANOVA test and to evaluate the relationship between the variables together, correlation test was used.

RESULTS AND DISCUSSION

Many studies in various species have been made regarding the influence zinc has on skin, in acceleration of wound healing (Pories et al., 1967; Senapati et al., 1990) and a part of a cellular differentiation process: the keratinization that transforms live epithelial cells into corneous cells that are structurally stable protective (Iwata et al., 1999) (Table 1-4 and Fig. 1 and 2). Chesters and Petrie (1999) showed that zinc has influence on both cell replication and differentiation. Others have shown effect of treatment on skin problems such as alopecia, superficiel flaking of dried epidermis, poor healing of abrasion and recurrent infections with edema, parakeratosis, seborrhea sicca and crusting dermathosis (Harrington et al., 1973; Van den Broek and Thoday, 1986; Sousa et al., 1988). When choosing the zinc source to equine feed, absorption must be considered.

The absorption of zinc occurs primarily in the small intestine (Weigand and Kirchgessner, 1976; Hambidge *et al.*, 1998). Once absorbed in plasma, zinc is bound to and transported by albumin and transferrin (Chesters and Will, 1981; Duchateau *et al.*, 1981). Since transferrin also transports iron, excessive iron reduces zinc absorption and vice-versa. The intake of excessive amounts of iron induces a decrease of plasma zinc levels in ponies. The intake of iron must be >800-2000 mg kg⁻¹ feed/day/horse before effects are detectable (Lawrence *et al.*, 1987).

Table 1: Zn distribution in the samples studied in the Summer

Variable	No.	Mean	SEM	SD	Min	Max
Zn	90	111.42	1.75	16.65	73	140

Table 2: Zn distribution in the samples studied in the winter							
Variable	No.	Mean	SEM	SD	Min	Max	
Zn	96	43.52	0.98	9.62	30	72.22	

Table 3: Comparison of zinc in serum in two season

Season	No.	Mean±SD	F-value	p-value	t-test	<u>df</u>
Winter	96	43.52±9.62	27.32	0.000	33.74	140.55
Summer	90	111.42±16.65				

Table 4.	Comparison	of zinc	in comm	hu cov

Sex	No.	Mean±SD	F-value	p-value	t-test	df
Male	162	76.01±63.53	0.041	0.84	0.26	184
Female	24	78.24±37.67				

A similar reaction occurs with copper, as metallothionein absorbs both zinc and copper. In intestinal cells, metallothionein is capable of adjusting absorption of zinc by 15-40%. However, inadequate or excessive zinc intake can be harmful.

Zinc supplementation with 161 mg zinc kg⁻¹ of Dry Matter (DM) feed along with excessive amount of calcium 1.25% DM feed showed a statistically significant fall of zinc concentration in blood being 1.03 mg L⁻¹ before and 0.86 mg L⁻¹ after (Danek *et al.*, 1999). Crozier *et al.* (1997) found that hay containing the amount of zinc recommended by American National Research Council (1989) still may require supplementation with phosphor, sulfur, cupper and crude protein to optimize the zinc absorption. Oral zinc preparations contain zinc in different types of salts and chelates which in turn affects

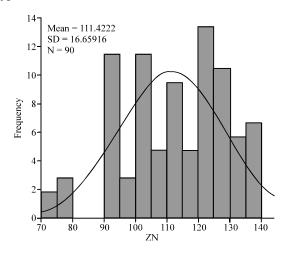


Fig. 1: Zn distribution in the samples studied in the Summer

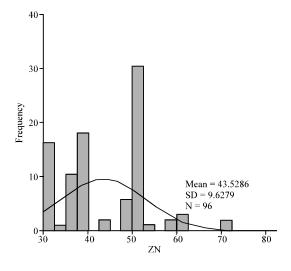


Fig. 2: Zn distribution in the samples studied in the Winter

the bioavailability of zinc. Bioavailability can be defined as a measurement of the rate and extent of a nutrient that reaches the systemic circulation and is available at target tissue level (Kienzle and Zorn, 2006). Wichert *et al.* (2002a, b) showed that the oral supplementation with zinc as zinc sulphate and zinc sulphate chelates to horses resulted in the highest increase in plasma zinc concentration compared to zinc oxide and zinc lactate.

The same tendency has been shown in chicks (Edwards and Baker, 1999; Wedekind et al., 1992). Although, an increase in serum zinc levels after long term oral supplementation of zinc oxide to ponies has been shown (Schryver et al., 1980), the tendency has later been shown to be of lower significance (Wichers et al., 2002). Lowe et al. (1994) showed that the rate of growth of hair and the amount of zinc deposited in the hair was significantly higher in dogs fed diets containing zinc as the amino acid chelate than in dogs fed zinc as zinc oxide or as a zinc polysaccharide complex. The 1989 NRC recommend that all classes of horses require 40 mg zinc kg-1 feed of dry matter per day. Jackson suggests the intake to be 400 mg Zn day⁻¹ for horses at light work and 500 mg day⁻¹ for a horse at moderate or heavy work, due to the zinc loss in sweat in working horses that Meyer found in 1986 (20-21 mg zinc L^{-1} of sweat).

Meyer et al. (2002) found that zinc requirement seems to be higher if the diet contains high levels of phytate, calcium or copper. When the diet is supplemented with as high as 1000 or 2000 mg zinc kg⁻¹ feed the copper metabolism is affected adversely in foals (Bridges and Muffit, 1990). The ratio of zinc to copper should not exceed 4:1 to 5:1 (Cymbaluk et al., 1986). Distal limb skin disease is a common debilitating problem in horses and to know whether the zinc status is below or above the normal range is essential in planning of treatment. This pilot study was conducted in order to observe the clinical effect of 2 months of oral organic zinc compound supplementation to seven horses with distal limb skin disease. Furthermore to observe the distribution of zinc in serum, hair and skin before and after oral zinc supplementation to horses with distal limb skin disease and hence to evaluate the paraclinical methods used to estimate equine zinc status.

Evaluation of the clinical effect is initially straightforward. There is sufficient improvement in all the horses, to validate the results. However, a control groups would have provided stronger evidence. It would exclude the possibility that the improvement could be due to weather, seasons, breed, age or gender. Assessment of the clinical results would quite evidently be stronger if the data material was constituted by a larger number. It would be beneficial to design more specific inclusion criteria

about etiologies of the disease and more standardized feeding before and during the study. Regarding SEM-EDS analysis of hair, it's easy to collect mane hair samples and it is without major inconvenience to the horse. When it comes to hair from the affected area many of the horses were fairly annoyed at the collection and precautions to avoid being kicked had to be taken.

Since, there also had to be taken biopsies, there was made perineural nerve block. The photo of magnifications of hair shows clearly the improvements. Regarding SEM-EDS analysis of skin and especially looking at the histopatological examination, it requires more in depth knowledge and experience to interpret. A good laboratory with experience in the field is recommended. The histopatological findings in this study match closely with the results of Sanecki et al. (1985) in puppies with skin problems related to zinc deficiency. Collection of the skin biopsy is more laborious and yields are not entirely worth the effort. The results of this study showed however as previous studies (Pories et al., 1967; Harrington et al., 1973; Van den Broek and Thoday, 1986; Sousa et al., 1988; Senepati, 1990; Iwata et al., 1999; Chester and Petrie, 1999; Rostan et al., 2002) that zinc therapy promotes healing and improves coat health.

Looking at Table 5, the positive effect is easy to spot. Assessment of the different paraclinical techniques to estimate the nutritional zinc status in the horses of this study has been limited by the lack of patients. It makes no sense to perform statistical calculations; they would at best be able to point to trends more than any significant evidence. However, the current results are of interest for further study. Regarding zinc serum concentration, it's easy to collect blood samples and it is without major inconvenience to the horse. The analysis of serum zinc concentrations of serum can be performed in a local laboratory and is not very time consuming. This paraclinical parameter is the most investigated and normal values are recognized to be applicable (National Research Council, 1989; SCAN, 2003).

The results must however be evaluated with caution since many variables come into play when estimating zinc concentration. The fact that all the horses were in the lower range of the reference values both before and after the treatment could be due to the fact that they were chronic infected or vice-versa, the low zinc concentration leading to chronic infection. Others had shown correlation between infection and serum concentration. Dede et al. (2008) shoved a decrease in plasma zinc in horses with piroplasmose and a correlation between a decrease in plasma zinc levels and increased activity of carbonic anhydrates. Carbonic anhydrates is the zinc containing enzyme that regulates the homeostasis of erythrocytes (Mafra and Cozzolino, 2004). Located in the lower end of the reference values or below, the diseased horse can certainly benefit from organic zinc supplements. Attempts to achieve a more accurate reflection of the tissue zinc and hence a precise estimate of horse zinc status using blood samples is done by Milne et al. (1985). They described a method for cell separation and analysis of separated platelets, mononucleated cell, polymorphnucleated cell and erythrocytes and measured individual zinc levels in each cell. Magneson et al. (1987) described the enzyme phosphoglucomutase as an indicator to measure free zinc in equine plasma. However, both these two methods are not accessible at laboratories.

CONCLUSION

In this study, results showed that there are significant differences in serum zinc levels in Summer and Winter.

REFERENCES

- Brandt, E.G., M. Hellgren, T. Brinck, T. Bergman and O. Edholm, 2009. Molecular dynamics study of zinc binding to cysteines in a peptide mimic of the alcohol dehydrogenase structural zinc site. Phys. Chem. Chem. Phys., 11: 975-983.
- Bridges, C.H. and P.G. Moffitt, 1990. Influence of variable content of dietary zinc on copper metabolism of weanling foals. Am. J. Vet. Res., 51: 275-280.
- Chesters, J.K. and L. Petrie, 1999. A possible role for cyclins in the zinc requirements during G1 and G2 phases of the cell cycle. J. Nutr. Biochem., 10: 279-290.
- Chesters, J.K. and M. Will, 1981. Zinc transport proteins in plasma. Br. J. Nutr., 46: 111-118.
- Crozier, J.A., V.G. Allen, N.E. Jack, J.P. Fontenot and M.A. Cochran, 1997. Digestibility, apparent mineral absorption and voluntary intake by horses fed alfalfa, tall fescue and caucasian bluestem. J. Anim. Sci., 75: 1651-1658.
- Cymbaluk, N.F., F.M. Bristol and D.A. Christensen, 1986. Influence of age and breed of equid on plasma copper and zinc concentrations. Am. J. Vet. Res., 47: 192-195.
- Danek, J., M. Gehrke and W. Krumrych, 1999. Effect of supplemental dietary zinc on hair zinc and blood serum zinc levels in stallions. Arbeitstagung Mengen- und Spurenelemente, pp. 768-775.
- Dede, S., Y. Deger, S. Deger and P. Tanritanir, 2008. Plasma levels of zinc, copper, copper/zinc ratio and activity of carbonic anhydrase in equine piroplasmosis. Biol. Trace Elem. Res., 125: 41-45.

- Duchateau, J., G. Delespesse and P. Vereecke, 1981. Influence of oral zinc supplementation on the lymphocyte response to mitogens of normal subjects. Am. J. Clin. Nutr., 34: 88-93.
- Edwards, H.M. and D.H. Baker, 1999. Bioavailability of zinc in several sources of zinc oxide, zinc sulfate and zinc metal. J. Anim. Sci., 77: 2730-2735.
- Hambidge, K.M., N.F. Krebs and L. Miller, 1998. Evaluation of zinc metabolism with use of stableisotope techniques: Implications for the assessment of zinc status. Am. J. Clin. Nutr., 68: 410S-413S.
- Harrington, D. D., J. Walsh and V. White, 1973. Clinical and pathological findings in the horse fed zincdeficient diets. Proc. 3rd Equine Nutr. Physiol., 1973: 51-54.
- Iwata, M., T. Takebayashi, H. Ohta, R.E. Alcalde, Y. Itano and T. Matsumura, 1999. Zinc accumulation and metallothionein gene expression in the proliferating epidermis during wound healing in mouse skin. Histochem. Cell Biol., 112: 283-290.
- Kienzle, E. and N. Zorn, 2006. Bioavailability of minerals in the horse. Proceedings of the 3rd European Equine Nutrition and Health Congress, March 17-18, 2006, Ghent University, Merelbeke, Belgium.
- Klevay, L.M., B.R. Bistrian, C.R. Fleming and C.G. Neumann, 1987. Hair analysis in clinical and experimental medicine. Am. J. Clin. Nutr., 46: 233-236.
- Lawrence, L.A., E.A. Ott, R.L. Asquith and G.J. Miller, 1987. Influence of dietary iron on growth, tissue mineral composition, apparent phosphorus absorption and chemical properties of bone. Proceedings of the 10th Equine Nutrition Physiological Society Symposium, June 11-13, 1987, Fort Collins, Colorado State University, pp. 563.
- Lowe, J.A., J. Wiseman and D.J.A. Cole, 1994. Zinc source influences zinc retention in hair and hair growth in the dog. J. Nutr., 124: 2575S-2576S.
- Mafra, D. and S.M.F. Cozzolino, 2004. Erythrocyte zinc and carbonic anhydrase levels in nondialyzed chronic kidney disease patients. Clin. Biochem., 37: 67-71.
- Magneson, G.R., J.M. Puvathingal and W.J. Ray Jr., 1987. The concentrations of free Mg2+ and free Zn2+ in equine blood plasma. J. Biol. Chem., 262: 11140-11148.
- McCarthy, T.J., J.J. Zeelie and D.J. Krause, 1992. The antimicrobial action of zinc ion/antioxidant combinations. J. Clin. Pharm. Ther., 17: 51-54.
- Milne, D.B., N.V. Ralston and J.C. Wallwork, 1985. Zinc content of cellular components of blood: Methods for cell separation and analysis evaluated. Clin. Chem., 31: 65-69.
- National Research Council, 1989. Nutrient requirements of horses. 5th Edn., National Academy of Sciences, Washington, D.C., pages: 100.

- Pories, W.J., J.H. Henzel, C.G. Rob and W.H. Strain, 1967. Acceleration of wound healing in man with zinc sulphate given by mouth. Lancet, 289: 121-124.
- Rostan, E.F., H.V. DeBuys, D.L. Madey and S.R. Pinnell, 2002. Evidence supporting zinc as an important antioxidant for skin. Int. J. Dermatol., 41: 606-611.
- SCAN., 2003. Opinion of the Scientific Committee for Animal Nutritionon the use of zinc in feedingstuffs. European Commissionhealth and Consumer Protection Directorate-General, http://ec.europa.eu/food/fs/sc/scan/out120 en.pdf.
- Sanecki, R.K., J.E. Corbin and R.M. Forbes, 1985. Extracutaneous histologic changes accompanying zinc deficiency in pups. Am. J. Vet. Res., 46: 2120-2123.
- Schryver, H.F., H.F. Hintz and J.E. Lowe, 1980. Absorption, excretion and tissue distribution of stable zinc and 65zinc in ponies. J. Anim. Sci., 51: 896-902.
- Senapati, A., B.M. Slavin and R.P. Thompson, 1990. Zinc depletion and complications of surgery. Clin. Nutr., 9: 341-346.

- Sousa, C.A., A.A. Stannard, P.J. Ihrke, S.I. Reinke and L.P. Schmeitzel, 1988. Dermatosis associated with feeding generic dog food: 13 cases (1981-1982). J. Am. Vet. Med. Assoc., 192: 676-680.
- Van den Broek, A.H.M. and K.L. Thoday, 1986. Skin disease in dogs associated with zinc deficiency: A report of five cases. J. Small Animal Prac., 27: 313-323.
- Wedekind, K.J., A.E. Hortin and D.H. Baker, 1992. Methodology for assessing zinc bioavailability: Efficacy estimates for zinc-methionine, zinc sulfate and zinc oxide. J. Anim. Sci., 70: 178-187.
- Weigand, E. and M. Kirchgessner, 1976. Radioisotope dilution technique for determination of zinc absorption in vivo. Nutr. Metab., 20: 307-313.
- Wichert, B., K. Kreyenberg and E. Kienzle, 2002a. Serum response after oral supplementation of different zinc compounds in horses. J. Nutr., 132: 1769S-1770S.
- Wichert, B., T. Frank and E. Kienzle, 2002b. Zinc, copper and selenium intake and status of horses in Bavaria. Am. Soc. Nutr. Sci., 132: 1776s-1777s.