Journal of Animal and Veterinary Advances 10 (24): 3244-3248, 2011

ISSN: 1680-5593

© Medwell Journals, 2011

Effects of *Hypericum perforatum* (St. John's wort) Extract on Renal Ischemic Reperfusion Injury in Rats

¹Ali Akbar Abolfathi, ²Yousef Doustar, ⁴Pejman Mortazavi and ³Ali Rezai ¹Deparetment of Pathobiology, Islamic Azad University, Ahar Branch, Ahar, Iran ²Department of Pathobiology, ³Department of Clinical Sciences, Islamic Azad University, Tabriz Branch, Tabriz, Iran ⁴Department of Pathology, Faculty of Specialized Veterinary Science, Science and Research Branch, Islamic Azad University, Tabriz, Iran

Abstract: Ischemia/Reperfusion (IR) induces severe tissue injury mainly caused by oxidative stress. A considerable body of recent evidence suggests that oxidative stress and exaggerated production of reactive oxidant species play a major role in several aspects ischemia and reperfusion. *Hypericum perforatum* is a medicinal plant species containing many polyphenolic compounds namely flavonoids and phenolic acids. Because polyphenolic compounds have high antioxidant potential in this study we evaluated the effect of H. perforatum extract on renal ischemic reperfusion injury. Renal ischemic reperfusion was induced in rats by clamping the renal artery for 60 min. After 7 days of reperfusion. *H. perforatum* extract treatment markedly reduced renal ischemia reperfusion injury in I/R+HP 30 mg kg⁻¹ group comparative with other treatment and sham groups.

Key words: Renal Ischemia/Reperfusion (IR), Hypericum perforatum, acute renal failure, tissue, Iran

INTRODUCTION

I/R induced Acute Renal Failure (ARF) is a common clinical problem which despite significant advances in critical care medicine is still associated with high morbidity and mortality. The mechanisms underlying renal I/R are complex including ATP depletion, accumulation of intracellular Ca2+ and reactive oxygen species, mitochondrial dysfunction, multiple enzyme systems activation and pro-inflammatory cytokine production. Although, reperfusion is essential for the survival of ischaemic renal tissue, it causes additional cellular injury. Together, renal ischaemia and reperfusion initiate a multiple and interrelated sequence of events resulting in the injury and eventually the death of renal cells as a of both apoptosis combination and (Spandou et al., 2009). Hypericum perforatum L. (Hypericaceae), popularly called St. John's wort has been used in popular medicine since ancient times for several disorders such as skin wounds, eczema, burns, diseases of the alimentary tract, insomnia and mental illness among others (El-Sherbiny et al., 2003). H. perforatum extract contains flavonoids such as rutin, quercetin and quercitrin which demonstrate a free radical scavenging activity in a model of auto-oxidation of rat cerebral membrances (Gambarana et al., 2001; Zou et al., 2004). An antioxidant activity of quercetin has also been

demonstrated by inhibition of brain lipid peroxidation as manifested by lowering MDA while elevating phospholipid contents in a rat model of endotoxemia (Benedi et al., 2004; Sanchez-Reus et al., 2007; Silva et al., 2005). Therefore, hypericum extract with a potential antioxidant activity may be of value in dementia among other disorders of senility in which free radical generation is implicated. In addition its antidepressant activities, H. peforatum in line with popular credence also possesses antiviral, wound-healing, antimicrobial, analgesic and anti-inflammatory effects (El-Sherbiny et al., 2003). Studies with other plants of the same genus have been carried out under the stimulus of great scientific interest and economic value acquired by H. perforatum. Thus, antidepressant, analgesic, anti-inflammatory, antioxidant, antimicrobial and wound-healing effects have also been found for other species of the genus Hypericum (Gambarana et al., 2001; Kerb et al., 1996). More recently, H. perforatum extract has been reported to efficiently attenuate Interferon (IFN)-v-elicited activation of Signal Transducers and Activators of Transcription (STAT)-1 in alveolar A549/8 and colon DLD-l cells (Tedeschi et al., 2003). Hypericum extract has been always referred to have a benign side effect profile compared with tricyclic antidepressants and serotonin-specific reuptake inhibitors (Nathan, 2001). There has not been a single fatal intoxication of the extract as a monotherapy reported in the literature (Kerb et al., 1996). Therefore, Hypericum extract as an efficacious antidepressant medication with a benign side effect profile together with a potential antioxidant activity was hypothesized to be useful in the treatment of pathological situation in which ROS play an important role such us ischemia and reperfusion. Thus, the aim of the present study was to evaluate the effect of *H. perforatum* extract on renal ischemic reperfusion.

MATERIALS AND METHODS

Animals: Studies were performed on male Wistar rats weighing 200-250 g (n = 25). Rats received a standard diet and water *ad libitum* and were housed in a 12 h light/dark cycle. The animals were randomly allocated into three groups: I/R-saline group in which rats were subjected to renal ischemia for 60 mm (n = 10); IR-*H. perforatum* (HP) group in which rats were administered HP (30 mg kg $^{-1}$ and gavages) 7 days prior to I/R and 7 day IR (n = 10); I/R-HP group in which rats were administered HP (60 mg kg $^{-1}$ and gavages) 7 days prior to I/R and 7 days after IR (n = 10), sham-operated group in which rats were subjected to identical surgical procedure without occlusion of both renal pedicles and maintained under anesthesia for the duration of the experiment (n = 5) (Bosco and Schweizer, 1988.

Surgical procedures: Animals were anesthetized with Ketamine hydrochloride (Ketamine = 10%, Alfasan, Woerden-Holland = 50 mg kg⁻¹) and Xylazine (Xylazin = 2%, Alfasan, Worden-Holland = 5 mg kg⁻¹) intraperitoneally. A midline incision was made in each rat and the left kidney became available. Ischemia-reperfusion injury was induced by applying a noncrushing microvascular clamp on the left renal artery for 60 min. After 60 min of ischemia, the clamp was removed and the tissue was closed in layers. The animals were then returned to their cages, reperfusion period was 7 days after surgery. In experimental group, 7 days before ischemia, HP 30 and 60 mg kg⁻¹ were administrated as orally form. Sham-operated animals underwent the same operation but without clamping. Animals were sacrificed after 7 days postoperatively under general anesthesia with an injection of over dosage of Thiopental sodium (60 mg kg⁻¹) and the left kidney was harvested.

Histopathology analyses: Kidneys sunk in 10% formalin buffer during 7 days for fixation then each of those segments were included in paraffin blocks, all specimens were serially sectioned longitudinally at 5 μm intervals and stained with Hematoxylin-Eosin (H and E) method and used for light microscopic examination under a Nikon microscope (ECLIPSE E200, Japan).

Extract preparation: Powdered leaves of HP were extracted 3 times with 1 L of 70% methanol (MeOH)/H₂O while being macerated at room temperature for 24 h each time. The hydroalcoholic extracts were combined and concentrated in vacuo to yield dried extract and it was defined by the producer as containing 0.28% hypericin. This hydroalcoholic extract was kept in refrigerator for all experiments.

RESULTS AND DISCUSSION

Histopathological changes of kidney in IR rats and IR rats pretreated with HP (30 and 60 mg kg⁻¹) have been shown in Fig. 1-3. In renal tissue of IR rats, acute tubular lytic necrosis, interstitial lymphocyte infiltration, hyperemia and wide hemorrhages were observed. Dilation of urinary space with hyaline proteinaceouse droplets adhesiveness of visceral and parietal bowmann's capsule, mesenchial moderate hypercellularity with hyperemia and some time the necrosis of squamous cell in parietal wall of bowmann's capsule were also prominent. In addition, hyaline casts in some time with obstruction and dilation of tubules were visible. Pretreatment with HP at a dose of 30 mg kg⁻¹ caused considerable reversion of pathological changes. In this group, low pathological injuries consisting of petty hyperemia and slight edema in tubular cells were distinguished. Mild infiltration of lymphocytes in interstitial areas was distinguishable. Pretreatment with

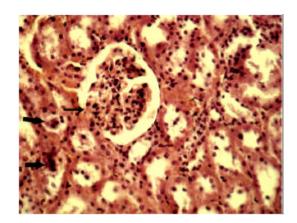


Fig. 1: Microscopic view of the kidney in I/R rat. Acute tubular lytic necrosis of the kidneys featuring detachment of the cells from the basal membrane, cytoplasmic granulation and pyknotic nuclei in both tubuli and glomeruli. Note the cellular debris in the lumens of these tubules (thick arrows). Lymphocyte infiltration in the interstitum is seen. Thickening of glomerular basement membrane, hypercellularity and synechiae (thin arrow) is also prominent (H and E stain 40x)

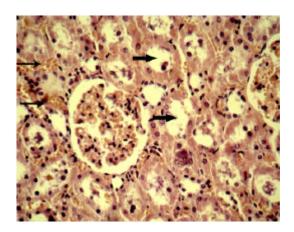


Fig. 2: Microscopic view of the kidney in I/R+HP 30 mg kg⁻¹ rat. Enlargement of lining cells of tubules with some tubular dilatation (thick arrows) is visible. There is mild hyperemia (thin arrows) and lymphocyte infiltration in the interstitial areas pathologic changes is mild to moderate comparison with representative sections from I/R rats (H and E stain, 40x)

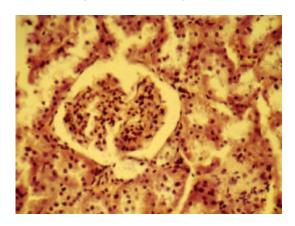


Fig. 3: Microscopic view of the kidney in I/R+HP 60 mg kg⁻¹ rat showing normal glomerulus and tubular cells. Pretreatment with HP (60 mg kg⁻¹) has reversed pathologic changes to near normalcy (H and E stain, 40x)

HP at a dose of 60 mg kg⁻¹ retrieved renal histopathological injuries to near normal. In Sham/IR group, the kidney tissue was normal and remarkably histological differences between them were not observed.

This study provides evidence that *H. perforatum* extract cases substantial reduction of renal ischemia/reperfusion injury in the rat. We speculate that *H. perforatum* extract and related compounds may be useful in the therapy of conditions associated with ischemica/reperfusion. Ischemia-reperfusion injury is a

major cause of renal failure and renal graft rejection. Therefore, reducing the extent of this injury in renal transplant patients is important for achieving a good prognosis. Renal Ischemia-Reperfusion (I/R) injury leads to the production of excess Reactive Oxygen Species (ROS) and Reactive Nitrogen Species (RNS). These species cause oxidative stress resulting in alterations in the level of mitochondrial oxidative phosphorylation, ATP depletion, increases in the intracellular calcium and activation of protein kinases, phosphatases, lipases and nucleases leading to a loss of cellular function and integrity (Sekhon et al., 2003). Therefore, it is important to reduce the levels of these hazardous methabolites in order to improve the patient's outcome. In order to reduce these metabolites, many studies have examined a variety of free radical oxygen scavengers. These include the effects of external supplementation of antioxidants (Gianello et al., 1996; Lee et al., 2006; Mathews and Gregory, 1997) and the activities of endogenous enzymatic antioxidant defense system in a kidney ischemia/reperfusion injury (Barnard et al., 1993; Dobashi et al., 2000).

In this study revealed that Hypericum perforatum (HP) caused decrease in renal tissue injury due to ischemic reperfusion so that was related to administrated dose of HP. Other researchers results also showed that low dose of Hypericum perforatum have more cytoprotective effect than mild doses 30 mg kg⁻¹ (Menegazzi et al., 2006). Hypericum perforatum is effective in treatment of diabetic patients and metabolic disorders (Menegazzi et al., 2006). Part of the therapeutic effects of Hypericum perforatum is related to antioxidant activation so that nowadays this drug is used in medical research as a stimulating of antioxidative stress pathway (El-Sherbiny et al., 2003). There are a few studies on effect of Hypericum perforatum on injuries subsequent ischemic reperfusion. De Paola et al. (2005) showed that short-term using of Hypericum perforatum during a mild ischemia improves cellular performance after ischemia on rats. Tedeschi et al. (2003), Hammer et al. (2007) showed that Hypericum perforatum have a anti-inflammatory reaction with inhibition of human inducible nitric oxide synthase expression by down regulating Signal Transducer and Activator of Transcription-1a (STAT-1) activation and Inhibition of Prostaglandin E2 Production. In all done studies about protective effect of Hypericum perforatum as single dose or low dose or short-term usage on injuries subsequent ischemic reperfusion and acute renal failure suggested that activation of antioxidative pathway is as Hypericum perforatum action mechanism (Flausino et al., 2002). Hunt et al. (2001) demonstrated that there is a free radical scavenging effect of H. perforatum extract and postulated that this effect could be related to an active

constituent. Likewise the beneficial effects of H. perforatum on acute cerebral ischemia-reperfusion could also be attributed to its bioactive constituents (Hunt et al., 2001). Benedi et al. (2004) and Sanchez-Reus et al. (2007) demonstrated that hypericum perforatum reduction of oxidative stress by inhibiting of free radical generation and lipid peroxidation. They also declared that Hypericum perforatum by activation of Superoxide Dismutase (SOD), Glutathione Peroxidase (GPx) and levels of Malondialdehyde (MDA) plays an important protective role on ischemic reperfusion injury (Benedi et al., 2004; Sanchez-Reus et al., 2007). Anyway, from this and other researches results can be conclude that Hypericum perforatum protects of renal ischemia reperfusion injury through inhibition of oxidative stress and inflammatory reactions occurrence in acute renal failure.

CONCLUSION

Considering the above-mentioned series can be declared that *Hypericum perforatum* can use as a preventive or harmless therapeutic agent against damages of renal ischemic reperfusion after controlled clinical trials. Thus to understanding of accurate sites and action mechanism of *Hypericum perforatum* requires many more expanded studies.

ACKNOWLEDGEMENT

Researcher thanks for Ahar Branch, Islamic Azad University for support.

REFERENCES

- Barnard, M.L., S.J. Snyder, T.D. Engerson and J.F. Turrens, 1993. Antioxidant enzyme status of ischemic and postischemic liver and ischemic kidney in rats. Free Radic. Biol. Med., 15: 227-232.
- Benedi, J., R. Arroyo, C. Romero, S. Martin-Aragon and A.M. Villar, 2004. Antioxidant properties and protective effects of a standardized extract of *Hypericum perforatum* on hydrogen peroxideinduced oxidative damage in PC12 cells. Life Sci., 75: 1263-1276.
- Bosco, P.J. and R.T. Schweizer, 1988. Use of oxygen radical scavengers on autografted pig kidneys after warm ischemia and 48-hour perfusion preservation. Arch. Surg., 123: 601-604.
- De Paola, R., C. Muia, E. Mazzon, T. Genovese and C. Crisafulli *et al.*, 2005. Effects of *Hypericum perforatum* extract in a rat model of ischemia and reperfusion injury. Shock, 24: 255-263.

- Dobashi, K., B. Ghosh, J.K. Orak, I. Singh and A.K. Singh, 2000. Kidney ischemia-reperfusion: Modulation of antioxidant defenses. Mol. Cell Biochem., 205: 1-11.
- El-Sherbiny, D.A., A.E. Khalifa, A.S. Attia, E.E.D.S. Eldenshary, 2003. Hypericum perforatum extract demonstrates antioxidant properties against elevated rat brain oxidative status induced by amnestic dose of scopolamine. Pharmacol. Biochem. Behav., 76: 525-533.
- Flausino, O.A., H. Zangrossi, J.V. Salgado and M.B. Viana, 2002. Effects of acute and chronic treatment with *Hypericum perforatum* L. (LI 160) on different anxiety-related responses in rats. Pharmacol. Biochem. Behav., 71: 251-257.
- Gambarana, C., P.L. Tolu, F. Masi, M. Rinaldi, D. Giachetti, P. Morazzoni and M.G. De Montis, 2001. A study of the antidepressant activity of *Hypericum perforatum* on animal models. Pharmacopsychiatry, 34: S42-S44.
- Gianello, P., A. Saliez, X. Bufkens, R. Pettinger, D. Misseleyn, S. Hori and B. Malfroy, 1996. EUK-134, a synthetic superoxide dismutase and catalase mimetic, protects rat kidneys from ischemiareperfusion-induced damage. Transplantation, 62: 1664-1666.
- Hammer, K.D.P., M.L. Hillwig, A.K.S. Solco, P.M. Dixon and K. Delate et al., 2007. Inhibition of prostaglandin E₂ production by anti-inflammatory Hypericum perforatum extracts and constituents in RAW264.7 mouse macrophage cells. J. Agric. Food Chem., 55: 7323-7331.
- Hunt, E.J., C.E. Lester, E.A. Lester and R.L. Tackett, 2001. Effect of St. John's wort on free radical production. Life Sci., 69: 181-190.
- Kerb, R., J. Brockmoller, B. Staffeldt, M. Ploch and I. Roots, 1996. Single-dose and steady state pharmacokinetics of hypericin and pseudohypericin. Antimicrob. Agents Chemother., 40: 2087-2093.
- Lee, J.I., M.J. Kim, C.S. Park and M.C. Kim, 2006. Influence of ascorbic acid on BUN, creatinine, resistive index in canine renal ischemia-reperfusion injury. J. Vet. Sci., 7: 79-81.
- Mathews, K.G. and C.R. Gregory, 1997. Renal transplants in cats: 66 cases (1987-1996). J. Am. Vet. Med. Assoc., 211: 1432-1436.
- Menegazzi, M., R. Di Paola, E. Mazzon, C. Muia and T. Genovese *et al.*, 2006. *Hypericum perforatum* attenuates the development of carrageenaninduced lung injury in mice. Free Radic. Biol. Med., 40: 740-753.
- Nathan, P.J., 2001. Hypericum perforatum (St John's Wort): A non-selective reuptakeinhibitor? A review of the recent advances in its pharmacology. J. Psychopharmacol., 15: 47-54.

- Sanchez-Reus, M.I., M.A. Gomez del Rio, I. Iglesias, M. Elorza, K. Slowing and J. Benedi, 2007. Standardized *Hypericum perforatum* reduces oxidative stress and increases gene expression of antioxidant enzymes on rotenone-exposed rats. Neuropharmacology, 52: 606-616.
- Sekhon, C.S., B.K. Sekhon, I. Singh, J.K. Orak and A.K. Singh, 2003. Attenuation of renal ischemia/reperfusion injury by a triple drug combination therapy. J. Nephrol., 16: 63-74.
- Silva, B.A., F. Ferreres, J.O. Malva and A.C.P. Dias, 2005. Phytochemical and antioxidant characterization of *Hypericum perforatum* alcoholic extracts. Food Chem., 90: 157-167.
- Spandou, E., I. Tsouchnikas, G. Karkavelas, E. Dounousi, C. Simeonidou, O. Guiba-Tziampiri and D. Tsakiris, 2009. Erythropoietin attenuates renal injury in experimental acute renal failure ischemic/reperfusion model. Nephrol. Dialysis Transplantation, 21: 330-336.
- Tedeschi, E., M. Menegazzi, D. Margotto, H. Suzuki, U. Forstermann and H. Kleinert, 2003. Antiinflammatory actions of St. John's wort: Inhibition of human inducible nitric-oxide synthase expression by down-regulating signal transducer and activator of transcription-1alpha (STAT-1alpha) activation. J. Pharmacol. Exp. Ther., 307: 254-261.
- Zou, Y., Y. Lu and D. Wei, 2004. Antioxidant activity of a flavonoid rich extract of *Hypericum perforatum L. in vitro*. J. Agric. Food Chem., 52: 5032-5039.