

## According to Canonical Correlation, the Evaluation of Bluefish (*Pomatomus saltatrix*) Blood Chemistry

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**Abstract:** Blood chemistry parameters can provide essential information on the physiological status of the animal and therefore allow accurate evaluations of the general health status. Canonical correlation analysis is a fundamental statistical tool. The goal of canonical correlation analysis is to evaluate the relative contribution of each variable to the derived canonical functions in order to explain nature of the relationships. CCA was used to determining, whether the blood protein parameters are related in any way to the blood lipids, enzymes, minerals. However, a linear association between predictor variables (blood proteins) and dependent variables (lipids, enzymes and minerals) were determining. These analyses results shown that canonical correlation analysis can be using prediction of relationships from blood proteins with other blood chemistry parameters.

**Key words:** Bluefish, *pomatomus saltatrix*, canonical correlation, blood chemistry, animals, enzymes

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### INTRODUCTION

The evaluation of blood chemistry parameters in animals is a routine and important tool in clinical practices. This simple technique can provide essential information on the physiological status of the animal and therefore, allow accurate evaluations of the general health status. However, the predictive value is compromised by the lack of reliable normal databases and available reference laboratories to properly analyze these samples (Berg and Bremset, 1998). The inference between these chemical measurements to certain diseases is mostly borrowed from experience in mammalian systems and only a few blood chemistry parameters have been confirmed experimentally, primarily in major fish species. In addition to our limited knowledge of blood chemistry of fish, the issue is further compounded by reports involving different sampling protocols as well as which parameters are determined. Moreover, many other factors including environmental (temperature, photoperiod, stock density, salinity) and physiological (reproductive cycle, age, gender, nutrition) have been reported to impact on blood parameters of fish. All of these have contributed to a limited use of blood chemistry parameters as a tool in fish health management.

It is well known that intensive fish culture is often accompanied by increased incidences of pathologies. Many studies have demonstrated the usefulness of hematology and blood biochemistry in the assessment of

fish health and as a biomarker of exposure to pollution (Bricknell *et al.*, 1999). Previous information on hematology and blood biochemistry in fish is fragmentary (Collazos *et al.*, 1998; Craig, 1977; De Pedro *et al.*, 1998). Exposures to environmental seasonal cycles in light, temperature and food availability are likely to affect blood and body composition. Indeed, seasonal changes in body composition, hematology and blood biochemistry have been described in several fish species (Groff and Zinkl, 1999; Handy and Depledge, 1999; Itazawa, 1957; Johnson and Wichern, 1988; Jonsson and Jonsson, 1998; Knoph and Masoval, 1996) but little is known to date on seasonal variations in body composition, hematology and blood biochemistry in fish.

Canonical Correlation Analysis (CCA) is a fundamental statistical tool. Canonical variables are linear combinations of the original quantitative measurements that contain the highest possible multiple correlation with each group and that summarize among-class variation (Leonard and McCormick, 1999).

The goal of CCA is to evaluate the relative contribution of each variable to the derived canonical functions in order to explain nature of the relationship(s). Consider the following two equations:

$$U_m = a_{m1}X_1 + a_{m2}X_2 + a_{mp}X_p \quad (1)$$

$$V_m = b_{m1}Y_1 + b_{m2}Y_2 + \dots + b_{mq}Y_q \quad (2)$$

Equation 1 and 2 gives the new variables  $U_m$  and  $V_m$ , which are a linear combination of the X (pre-slaughter) and Y (after slaughter) variables respectively. Let  $C_m$  be the correlation between  $U_m$  and  $V_m$ .

The objective of canonical correlation is to estimate  $a_{m1}, a_{m2}, \dots, a_{mq}$  and  $b_{m1}, b_{m2}, \dots, b_{mq}$  such that  $C_m$  is maximum. Equation 1 and 2 are the canonical equations,  $U_m$  and  $V_m$  are the canonical varieties and  $C_m$  is the canonical correlation (Master *et al.*, 1990).

In the study, CCA was used to determine whether the blood protein parameters are related in any way to the blood lipids, enzymes, minerals. From canonical correlation, a linear association between predictor variables (blood proteins) and dependent variables (lipids, enzymes and minerals) were determined.

## MATERIALS AND METHODS

Sample collection was monthly performing 3 different areas between December and February on Dardanelles (Fig. 1).

In each collection was caught 30 Bluefish (*Pomatomus saltatrix*) caught, weighed and measured. Mean Bluefish sizes were measured  $30.62 \pm 0.20$  cm and  $349.89 \pm 8.26$  g. Some chemical properties of sea water were shown in Table 1.

Immediately after capture fish were cleaned to prevent mucus contamination and blood samples were collected.

Blood was sampled from the caudal vein using a gauge needle and 5 mL syringe and then later transported laboratory. Chemical analyses were conducted using an enzymatic auto analyzer (Svoboda *et al.*, 2001; Val *et al.*, 1998; Rowley *et al.*, 1988).

Blood samples were centrifuged for 10 min at  $6.67 \times 10^{-8}$  g and the extracted serums were analyzed using ILab 900 and 1800 auto analyzer (Xia, 2008).

Biochemical parameters analyzed Urea (U), Creatine (CR), Uric Acid (UA), Total Protein (TP), Albumin (ALB), Globulin (GLB), Total Bilirubin (TBL), Direct Bilirubin (DBL), Indirect Bilirubin (IBL), Cholesterol (CHL), Triglycerides (TRG), High-Density Lipoproteins (HDL), Low-Density Lipoproteins (LDL), Very Low-Density Lipoproteins (VLDL), Alanine Amino Transferase (ALT), Aspartate amino Transferase (AST), Lactate Dehydrogenase (LDH), Alkaline Phosphatase (ALP), Amylase (AMY), Sodium (Na), Potassium (K), Chloride (Cl), Phosphorus (P), Iron (Fe) and Calcium (Ca). Statistical analyses were performed with SAS PROC CANCORR.

Table 1: Chemical properties of sea water on Dardanelles

Months	NO <sub>2</sub>	NO <sub>3</sub>	NH <sub>4</sub>	Temperature	
				(°C)	Salinity (%)
December	0.0007	0.108	0.002	11	22
January	0.0020	0.200	0.023	9.5	22
February	0.0032	0.106	0.08	8.5	23

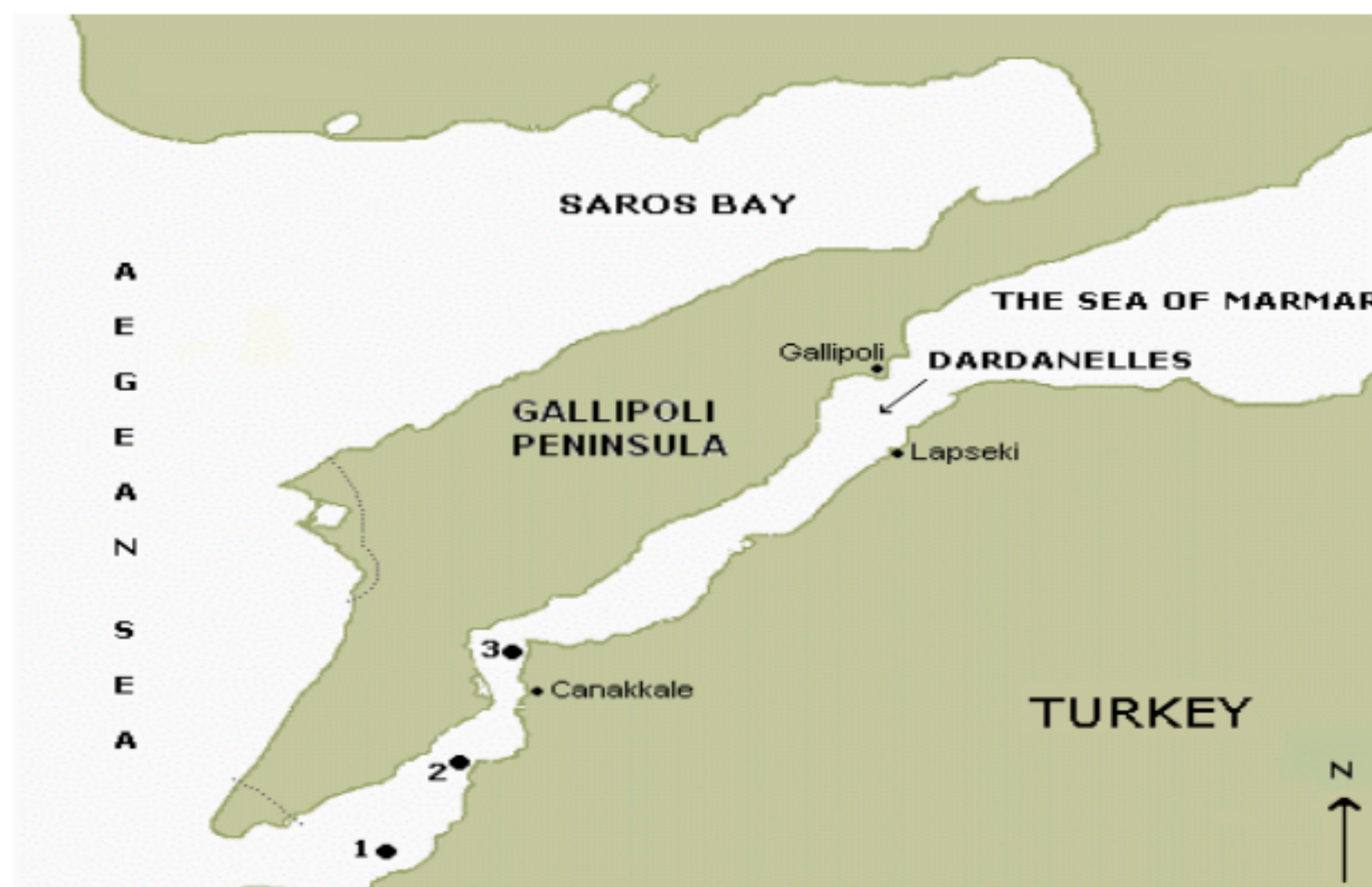


Fig. 1: Map of Dardanelles (1, 2, 3: represent stations where Bluefish caught)



## RESULTS

Table 2 shows the correlations among the blood proteins and lipids and enzymes (original variables). The correlations between the blood proteins and lipids parameters are moderate, the largest being 0.87 between TP and LDL and followed by TBL and CHL (0.82) and TBL and LDL (0.80).

The correlations between the blood proteins and enzymes are also moderate, the largest being 0.77 between DBL and ALP and 0.75 between IBL and alp. The correlations between the blood proteins and minerals are small, the largest being 0.54 between UA and Na, DBL and Na and DBL and Na (Table 3). Table 4 displays the correlations of blood lipids with enzymes and minerals.

The correlations between the blood lipids and enzymes are moderate, the largest being -0.90 between alt and Na followed by the correlation between ALP and Na (0.82). On the other hand, the correlations between the blood lipids and minerals are small, the largest being -0.57 between HDL and Na.

The correlations among the enzymes and minerals are moderate, the largest being -0.90 between alt and Na followed by the correlation between ALP and Na (Table 5). It was shown that the 0.997 calculated canonical correlation between blood proteins and blood lipids was significant ( $p < 0.01$ ). Relationship among the blood protein parameters of bluefish were expressed as following and canonical correlation was shown in Table 6.

Table 2: Correlations among the blood proteins and lipids and enzymes

Proteins	Lipid/enzymes									
	CHL	TRG	HDL	VLDL	LDL	AST	ALP	LDH	ALT	AMY
U	0.17	0.30	-0.43*	0.30	0.33	0.52*	0.38	-0.13	-0.37	-0.25
CR	0.15	-0.02	-0.37	-0.01	0.09	0.06	0.07	-0.41	-0.11	-0.44*
UA	0.55**	-0.07	-0.32	-0.07	0.61*	0.38	0.50*	-0.22	-0.40*	-0.33
TP	0.72**	0.53**	-0.58**	0.52**	0.87**	0.29	0.43*	-0.55*	-0.26	-0.30
ALB	0.40*	0.36	-0.59**	0.36	0.62**	0.44*	0.50*	-0.58*	-0.33	0.50*
TBL	0.82**	0.59**	-0.32	0.58**	0.80**	0.05	-0.01	-0.27	0.01	-0.08
DBL	0.08	0.02	-0.47*	0.02	0.23	0.53*	0.77**	-0.43	-0.38	-0.53*
IBL	0.11	0.32	-0.55**	0.32	0.32	0.57*	0.75**	-0.49	-0.42*	-0.53*

Table 3: Pearson-moment correlations among the blood proteins and minerals

Proteins	Minerals					
	Na	K	Cl	P	Fe	Ca
U	0.31	0.19	0.20	0.07	-0.04	0.25
CR	0.26	0.25	0.10	-0.10	0.07	0.17
UA	0.54**	0.37	0.41*	0.37	0.15	0.49
TP	0.44*	0.30	0.33	0.40*	0.43*	0.48*
ALB	0.48*	0.26	0.20	0.29	0.34	0.46*
TBL	0.05	0.19	0.25	0.30	0.42*	0.29
DBL	0.54**	0.03	0.23	0.25	0.11	0.42*
IBL	0.54**	0.09	0.27	0.29	0.24	0.48*

Table 4: Pearson-moment correlations among the blood lipids and enzymes and minerals

Lipids	Enzyme/minerals										
	AST	ALP	LDH	ALT	AMY	Na	K	Cl	P	Fe	Ca
CHL	-0.27	-0.17	-0.39	0.18	0.12	-0.07	0.14	0.14	0.15	0.28	0.10
TRG	0.13	-0.03	-0.33	-0.05	0.05	-0.03	0.10	0.06	0.18	0.31	0.16
HDL	-0.64**	-0.60**	0.41	0.53**	0.42*	-0.57*	-0.33	-0.25	-0.28	-0.19	-0.41*
VLDL	0.14	-0.01	-0.33	-0.04	0.04	-0.03	0.09	0.06	0.17	0.30	0.15
LDL	0.17	0.18	-0.43	-0.24	-0.12	0.33	0.43*	0.37	0.39	0.38	0.41*

Table 5: Pearson-moment correlations among the blood enzymes and minerals

Enzyme	Minerals					
	Na	K	Cl	P	Fe	Ca
AST	0.69**	0.55**	0.39	0.40	-0.02	0.43*
ALP	0.82**	0.30	0.33	0.31	-0.09	0.49*
LDH	-0.39	-0.25	0.04	0.01	0.04	-0.14
ALT	-0.90**	-0.70**	-0.65*	-0.52**	-0.25	-0.66**
AMY	-0.68**	-0.25	-0.24	-0.10	0.11	-0.35

\* $p < 0.05$ , \*\* $p < 0.01$

Table 6: Canonical Correlation Coefficients, R<sup>2</sup> and p-values

C.Cr.	R <sup>2</sup>	p-values	Canonical varieties
<b>Proteins: Lipids</b>			
0.997	0.994	0.00	V <sub>1</sub> =0.4788X1+0.4495X2+0.3802X3+1.5272X4-0.1300X5-1.1109X6+0.2057X7-0.8064X8-0.5642X9 W <sub>1</sub> =0.2397Y1-18.8105Y2-0.9316Y3+18.3536Y4+0.3796
<b>Proteins: Enzymes</b>			
0.998	0.996	0.00	V <sub>1</sub> =-8.784X1-4.109X2+12.32X3-21.33X4+17.9X5+1.56X6+0X7+0X8+0X9 W <sub>1</sub> =-0.457Y1+1.20Y2+1.22Y3-0.481Y4+0.252Y5
<b>Proteins: Minerals</b>			
0.989	0.979	0.00	V <sub>1</sub> =0.3168X1+0.3618X2-0.4957X3+1.4799X4-1.087X5+0.2438X6-0.1714X7-0.5541X8+0.1853X9 W <sub>1</sub> =-1.0305Y1+1.1741Y2+1.0265Y3-0.2967Y4+0.2515Y5-0.6448Y6
<b>Lipids: Enzymes</b>			
0.246	0.06	0.55	V <sub>1</sub> =-0.58X1-21.52X2+4.45X3+22.86X4+3.26X5 W <sub>1</sub> =-1.35Y1+1.91Y2-0.61Y3-0.06Y4+0.45Y5
<b>Lipids: Minerals</b>			
0.955	0.914	0.00	V <sub>1</sub> =-0.69X1-2.46X2+0.14X3+2.57X4+1.56X5 W <sub>1</sub> =0.23Y1+0.83Y2-0.46Y3+0.13Y4+0.25Y5+0.22Y6
<b>Enzymes: Minerals</b>			
0.76	0.578	0.096	V <sub>1</sub> =-2.01X1+0.82X2-0.95X3+0.81X4-1.67X5 W <sub>1</sub> =-0.13Y1+1.87Y2-14.01Y3+0.29Y4+4.05Y5+8.92Y6

$$V_1 = 0.4788U + 0.4495CR + 0.3802UA + 1.5272TP - 0.1300ALB - 1.1109GLB + 0.2057TBL - 0.8064DBL - 0.5642IBL$$

While relationship among the blood lipid parameters were expressed as following equation:

$$W_1 = 0.2397CHL - 18.8105TRG - 0.9316HDL + 18.3536VLDL + 0.3796LDL$$

## DISCUSSION

Evaluation of the relationship between these parameters the above equations was sufficient. R<sup>2</sup> value of the canonical correlation is 99.4%. When coefficients of the V<sub>1</sub> and W<sub>1</sub> equations were examined, it was seen that while CHL, VLDL and LDL lipids were affected as positively from U, CR, UA, TP and TBL, while TRG and HDL were negatively affected. It can be said that TP with a contribution of 1.5272 is the most determinative protein in the increase in CHL, VLDL and LDL lipids. On the other hand, the cholesterol, VLDL and LDL lipids were affected as negatively from ALB, GLB, DBL and IBL proteins. These results suggested that, those bluefish with the higher U, CR, UA, TP and TBL tend to also have higher CHL, VLDL and LDL, while they tend to low TRG and LDH. Likewise, those bluefish with the higher ALB, GLB, DBL and IBL also have TRG and HDL.

It was shown that the 0.998 calculated canonical correlation between blood proteins and blood enzyme was significant (p<0.01). R<sup>2</sup> value of the canonical correlation is 99.6%. When coefficients of the V<sub>1</sub> and W<sub>1</sub> canonical varieties were examined, it was seen that while ALT, LDH and AMY enzymes were affected as positively from UA,

ALB and GLB, AST and ALP were negatively affected. On the other hand, TBL, DBL and IBL had no effect on any of the blood enzymes measured.

These results suggested that those bluefish with the higher UA, ALB and GLB tend to also have higher ALT, LDH and AMY, while they tend to low AST and ALP. Likewise, those bluefish with the higher U, CR and TP also have AST and ALP. It can be show that UA with a contribution of 12.32 is the most determinative protein in the increase in ALT, LDH and AMY enzymes, while UA with a contribution of 21.33 is the most determinative protein in the increase in AST and ALP enzymes.

It was shown that the 0.989 calculated canonical correlation between blood proteins and blood minerals was significant (p<0.01). R<sup>2</sup> value of the canonical correlation is 97.9%. When coefficients of the V<sub>1</sub> and W<sub>1</sub> canonical varieties were examined, it was seen that while Na, P and Ca enzymes were affected negatively from U, CR, TP, GLB and IBL, K, Cl and Fe were positively affected. These results suggested that those Bluefish with the higher U, CR, TP, GLB and IBL tend to also have lower Na, P and Ca while they tend to higher K, Cl and Fe minerals. At the same time, those bluefish with the higher UA, ALB, TBL and DBL also have Na, P and Ca minerals. It can be show that ALB with a contribution of 1.087 is the most determinative protein in the increase in Na, P and Ca minerals.

Canonical correlation coefficients for lipids-enzymes and enzymes-minerals were 0.246 (p = 0.55) and 0.76 (p = 0.096), respectively. However, the canonical correlation coefficient for lipids-minerals was 0.955 (p<0.01). R<sup>2</sup> value of the canonical correlation is 91.4%. When coefficients of the V<sub>1</sub> and W<sub>1</sub> canonical varieties were examined, it was seen that while Na, K, P, Fe and Ca

minerals were affected as negatively from CHL and trig or vice versa, Cl was positively affected. These results suggested that those bluefish with the higher CHL and TRG tend to also have lower Na, K, P, Fe and Ca minerals, while they tend to have higher Cl. At the same time, those Bluefish with the higher HDL, VLDL and LDL also have Na, K, P, Fe and Ca minerals. It can be said that VLDL with a contribution of 2.57 is the most determinative lipids in the increase in Na, K, P, Fe and Ca minerals.

## CONCLUSION

As results of these analyses, CCA was used to determining whether the blood protein parameters are related in any way to the blood lipids, enzymes, minerals. However, a linear association between predictor variables (blood proteins) and dependent variables (lipids, enzymes and minerals) were determining.

## REFERENCES

- Berg, O.K. and G. Bremset, 1998. Seasonal changes in the body composition of young riverine Atlantic salmon and brown trout. *J. Fish Biol.*, 52: 1272-1288.
- Bricknell, I.R., T.J. Bowden, D.W. Bruno, P. MacLachlan, R. Johnstone and A.E. Ellis, 1999. Susceptibility of atlantic halibut, *Hippoglossus hippoglossus* (L.) to infection with typical and atypical *Aeromonas salmonicida*. *Aquaculture*, 175: 1-13.
- Collazos, M.E., E. Ortega, C. Barriga and B. Rodriguez, 1998. Seasonal variation in haematological parameters in male and female *Tinca tinca*. *Mol. Cell. Biochem.*, 183: 165-168.
- Craig, J.F., 1977. The body composition of adult perch, *Perca fluviatilis*, in Windermere, with reference to seasonal changes and reproduction. *J. Anim. Ecol.*, 46: 617-632.
- De Pedro, N., M.J. Delgado, M.L. Pinillos, A.L. Alonso-Gomez and M. Alonso-Bedate, 1998. Daily rhythms in NAT activity, cortisol, glucose, glycogen and catecholamines in tench (*Tinca tinca* (L.)). *Pol. Arch. Hydrobiol.*, 45: 321-329.
- Groff, J.M. and J.G. Zinkl, 1999. Hematology and clinical chemistry of cyprinid fish common carp and goldfish. *Vet. Clin. North Am. Exot. Anim. Pract.*, 2: 741-776.
- Handy, R.D. and M.H. Depledge, 1999. Physiological responses: Their measurement and use as environmental biomarkers in ecotoxicology. *Ecotoxicology*, 8: 329-349.
- Itazawa, Y., 1957. Gas content of the blood in response to that of medium water in fish. *Bull. Jap. Soc. Sci. Fish.*, 23: 71-80.
- Johnson, R.M. and D.W. Wichern, 1988. *Applied Multivariate Statistical Analysis*. Prentice Hall Int. Inc., Englewood Cliffs, NJ., USA., pp: 607.
- Jonsson, N. and B. Jonsson, 1998. Body composition and energy allocation in life-history stages of brown trout. *J. Fish Biol.*, 53: 1306-1316.
- Knoph, M.B. and K. Masoval, 1996. Plasma ammonia and urea levels in Atlantic salmon farmed in sea water. *J. Fish Biol.*, 49: 165-168.
- Leonard, J.B.K. and S.D. McCormick, 1999. Changes in haematology during upstream migration in American shad. *J. Fish Biol.*, 54: 1218-1230.
- Master, L.B.R., J.A. Brock, R.S. Fujioka and R.M. Nakamura, 1990. Hematologic and blood chemistry values for *Sarotherodon melanotheron* and a red hybrid tilapia in freshwater and seawater. *Comp. Biochem. Physiol.*, 97: 525-529.
- Rowley, A.F., T.C. Hunt, M. Page and G. Mainwaring, 1988. Fish. In: *Vertebrate Blood Cells*, Rowley, A.F. and N.A. Ratcliffe (Eds.). Cambridge University Press, Cambridge, pp: 19-127.
- Svoboda, M., J. Kouril, J. Hamackova P. Kalab, L. Savina, Z. Svobodova and B. Vykusova, 2001. Biochemical profile of blood plasma of tench (*Tinca tinca* L.) during pre-and postspawning period. *Acta Vet. Brno*, 70: 259-268.
- Val, A.L., G.C. de Menezes and C.M. Wood, 1998. Red blood cell adrenergic responses in *Amazonian teleost*. *J. Fish Biol.*, 52: 83-93.
- Xia, Y., 2008. A semiparametric approach to canonical analysis. *J. R. Statist. Soc. Series B: Statist. Methodol.*, 70: 519-543.